

ISSN Print: 2617-4693 ISSN Online: 2617-4707 IJABR 2024; 8(5): 828-840 www.biochemjournal.com Received: 13-03-2024 Accepted: 18-04-2024

Kratika Nayak

Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Rishikesh Tiwari Department of Soil Science and Agricultural Chemistry JNKVV, Jabalpur, Madhya Pradesh, India

Vinita Parte Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Monika Chouhan Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Pramod Kumar Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Badal Verma Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Renu Jayant Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Anamika Pandey Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Corresponding Author: Kratika Nayak Department of Agronomy, JNKVV, Jabalpur, Madhya Pradesh, India

Bio-herbicides: An eco-friendly approach for integrated weed management

Kratika Nayak, Rishikesh Tiwari, Vinita Parte, Monika Chouhan, Pramod Kumar, Badal Verma, Renu Jayant and Anamika Pandey

DOI: https://doi.org/10.33545/26174693.2024.v8.i5j.1201

Abstract

Bio-herbicides offer a promising eco-friendly alternative to chemical herbicides, addressing concerns over environmental impact and human health risks. These bio-controls, like Devine, Collego, and Biomal, are effective, economically feasible, and non-toxic to non-target organisms. They can significantly reduce weed-related crop yield losses, which account for 37% of overall pest-induced losses. However, the development of bio-herbicides requires careful consideration of formulation, fermentation systems, and strategic evaluation to ensure their effectiveness and commercial viability. To enhance the adoption and efficacy of bio-herbicides, researchers must address key constraints, including environmental, biological, technological, and commercial challenges. These constraints hinder the widespread use of bio-herbicides and limit their effectiveness as a sole weed management solution. While not a total replacement for chemical herbicides, bio-herbicides can serve as a vital component of integrated weed management systems, particularly as advancements in target selection, formulation, and marketing are made to overcome current limitations.

Keywords: Augmentative, bioherbicides, pathogen, commercialization, adjuvants and myco-herbicides

Introduction

Weeds pose significant challenges in agriculture by competing with crops for essential resources like water, nutrients, and light. They can also serve as hosts for pests and diseases, further impacting crop health and yield (Nichols, 2015) ^[129]. Weeds can cause significant yield losses in various crops, including: Direct-seeded rice: 15 to 66%, Maize: 18 to 65%, Soybean: 50 to 76% and Groundnut: 45 to 71% (Gharde, 2018) [56]. The extent of crop yield losses caused by weeds varies significantly and is influenced by several factors, including the type of crop, weed management approaches employed, weed species present, duration of weed infestation, and environmental factors such as climate and soil characteristics (Oerke et al., 2006) ^[131]. Weed management is a critical practice in agriculture. Due to labor shortages, there is a growing global trend towards increased use of herbicides to control weed populations (Aktar, 2009)^[4]. Inevitably, the constant use of herbicides on the same field to control weeds over a prolonged period has been shown to cause herbicide resistance, residue in crops, ecological imbalance between harmful and beneficial organisms, and environmental pollution. However, time constraint, advances in pest control technology, as well as a continuous 'enticement' from the current agricultural system have encouraged farmers to keep using conventional herbicides which have been found to be effective and time- and cost-efficient. The application of synthetic herbicides for effective weed control has thus become indispensable despite the unwelcome side effects. Organic fruits, vegetables, dairy products, and beverages have been increasingly popular worldwide, especially in developed nations. This trend reflects a growing consumer preference for organic food (Somasundram, 2016) ^[174]. Organic products represent a small portion of the food industry, yet their rapid expansion has sparked significant interest among consumers, businesses, and researchers alike. In 2013, there were nearly two million organic producers worldwide. Asia accounted for 36% of global organic farmers, followed by Africa at 29% and Europe at 17% (Somasundram, 2016)^[174]. Organic product sales have consistently increased over the last decades (Willer, 2015) ^[194]. To meet increasing consumer demand, farmers are transitioning from chemical-dependent conventional agriculture to sustainable, environmentally friendly

practices. This shift has led to the emergence of more sustainable weed control alternatives. Sustainable weed management focuses on preventing weed spread rather than controlling them after they have already become problematic. Sustainable weed management includes various practices like crop rotation, intercropping, crop competitiveness, tillage, mulching, biological control agents, and bioherbicides. These methods aim to control weeds without relying on chemical herbicides. Biological weed control involves using natural enemies, substances, or agents to limit weed populations to a manageable level economically. The application methods for bioherbicides are similar to conventional herbicides, but with mycoherbicides, the pathogenic fungi are sprayed onto target weeds to inoculate them. Recently, bio herbicides have been regarded as a crucial weed control element (Hoagland, 2007)^[65] albeit not as a total replacement but rather as an alternative to chemical herbicides (Singh, 2009 Raghuwanshi et al., 2023a; Yadav et al., 2023a, Singh et al., 2022; Jha et al., 2023, Kantwa et al., 2019; Yadav et al., 2023c; Singh et al., 2013b) [168, 145, 197, 85, 91, 199, 167]. Sustainable weed management is characterized by its reliance on a variety of weed control techniques, rather than a single approach, as seen with synthetic herbicides in conventional agriculture. Therefore, to effectively control weeds, bio-herbicides should be used in conjunction with other weed management methods (Yadav et al., 2023b; Patel et al., 2023, Singh et al., 2013a; Sahu et al., 2022) [198, 137, 164, 152]. Bio herbicides that are thought to be safer and 'greener' have drawn attention, as scientific reports provide increasing evidence of their efficacy. However, their commercial presence in comparison to conventional herbicides is relatively new. Therefore, rigorous testing and validation is necessary to evaluate their efficacy and reliability for weed control (Tiwari et al., 2011a; Tomar et al., 2023a, Kumar et al., 2022; Raghav et al., 2023; Tanisha et al., 2022) [186, 188-189, ^{104, 144, 179]}. This review examines the influence of bioherbicides and their limitations. The article aims to determine the role of bio-herbicides in weed control, including their incorporation into existing systems, to analyze factors influencing their effective use in both traditional and alternative management practices, to evaluate additional benefits beyond weed control, (Malviya et al., 2012, Sahu et al., 2022) ^[118, 152] and to assess the obstacles to the future advancement of bio-herbicides for efficient biologically-based weed management.

Bio herbicides

Bioherbicides consist of microorganisms such as pathogens and other microbes or phytotoxins derived from microbes, insects, or plant extracts that act as a natural means of weed control. According to Bailey, bioherbicides are naturally originated products which can be used to control weeds. But one must remember that, although bioherbicides comprise nature-derived compounds, this is not to say they are completely harmless. Plants naturally produce toxins that can impact the health of non-plant organisms in the environment. These toxins can also affect certain bacteria, viruses, and fungi, potentially leading to health issues in animals and humans (Sekhar, 2012) ^[162]. Thus, it is crucial to manage these natural toxins carefully to prevent unintended harm to crops or beneficial organisms in the environment (Duke, 2000) ^[49-50]. The first evidence of bioherbicide development was documented in the mid1970s with the discovery of mycoherbicides. Since then, numerous bioherbicides have been registered and become available in the global market (Zeng, 2020) ^[200]. The earliest bioherbicide project involved simply the application of *Fusarium oxysporum* Schlecht, a fungus, against *Opuntia ficus-indica* (L.) Mill, (Pacanoski, 2015) ^[134]. In the 1950s, the parasitic weed Cuscata spp. was controlled with Alternaria cuscutacidae Rudakov (Pacanoski, 2015) ^[134]. Meanwhile, the utilization of both registered and unregistered bio-herbicides has risen notably. In the late 1960s, an ambitious effort was made to discover pathogens from Rumex spp. in the United States (Inman, 1971) ^[70] and Rubus spp. in Chile (Oehrens, 1977) ^[130] for weed control.

Steps in developing a bioherbicide

Certainly! Here's a more detailed explanation of the three essential stages in the development of organic or natural herbicides:

Detection Stage

- a) Assembly of Unhealthy Flora: Researchers identify and collect plant pathogens or microbes that have the potential to act as herbicidal agents.
- **b)** Separation of Underlying Creatures: Once potential pathogens are collected, they are isolated from other microorganisms to ensure purity.
- c) Application of Koch's Hypotheses: Koch's postulates are applied to confirm that the isolated pathogen is the causative agent of disease in plants.
- d) **Pathogen Recognition:** The specific pathogen is identified and characterized for its herbicidal properties.
- e) Pathogen Cultivation on Synthetic Medium: The isolated pathogen is grown in a controlled synthetic medium to produce sufficient biomass for further studies.
- **f) Maintenance of Crops:** The cultivated crops are maintained for short-term or long-term storage, depending on the requirements of the research.

Phase of Progression

- a) Creating Suitable Conditions for Spore Growth: Optimal conditions are provided for the pathogen to produce spores, which are the infectious agents.
- b) Suitable Conditions for Contamination and Progression of Infection: Conditions that promote the spread of the pathogen and its infection of target weed species are identified and optimized.
- c) Assessing Host Range with Exposure to Pathogen Exploitation Appliances: The range of plant species that can be affected by the pathogen is determined through controlled experiments.

Stage of Placement

- a) Close Association Between Non-industrial and Developed Sectors: Collaboration between academia, research institutions, and industry is established to facilitate the development, field assessment, and commercialization of the bioherbicide.
- **b) Growth:** The production process is scaled up to produce the bioherbicide in larger quantities.
- c) **Field Assessment:** The efficacy and safety of the bioherbicide are tested in field trials under real-world conditions.

d) Publication of Phases of the Bioherbicide Production Marketing Cycle: The successful development and marketing of the bioherbicide are publicized to promote its adoption in sustainable agriculture practices.

Some commercially available bioherbicides

Certainly! Here's a detailed explanation of the bioherbicides we mentioned:

Phytophthora palmivora (Devine)

Developed by Abbott Laboratories, USA, it was the first mycoherbicide derived from the fungus *Phytophthora palmivora*. It targets Morrenia odorata, causing deadly root rot and collar rot. It can survive in the soil for extended periods. It was introduced in 1981 (Rao, 2000)^[147].

Colletotrichum gloeosporioides (Collego)

This mycoherbicide was developed from the anthracnose

fungus *Colletotrichum gloeosporioides* f. sp. *Aeschynomene*. It is effective against *Aeschynomene virginica* in rice and soybean crops. It was the first mycoherbicide commercially available for controlling annual wild plants in crops with over 90 percent efficacy. Named in 1982 (Boyette *et al.*, 2012)^[26].

Colletotrichum gloeosporioides (Biomal): Developed by Philom BiosInc., Canada, it is another mycoherbicide based on *Colletotrichum*. It targets *Malva pusilla*. It includes C-Spores. Successfully developed by Collego.

Streptomyces viridochromogenes (Bialaphos and Glufosinate): Bialaphos and Glufosinate are derived from Streptomyces viridochromogenes. It is a by-product obtained through fermentation. It is the active component of bialaphos, obtained from non-phytopathogenic Streptomyces. These bioherbicides are available internationally. (Carbonari et al. 2016)^[29].

Trade name	Pathogen	Weed host	Country	Reference
Collego®	Colletotrichum gloeosporioides f. sp. aeschynomene	Northern joint vetch	USA	Bowers, 1986 ^[22] ; Smith, 1982, 1991 ^{[171-} 172]
BioMal®	Colletotrichum gloeosporiodes f. sp. malvae	Round leaved mallow	USA; Canada	Boyetchko, 1998 ^[24] ; Mortensen, 1998 ^[126] ; Mortensen and Makowski, 1997 ^[125]
Hakatak™	Colletotrichum gloeosporiodes	Silky Hakea	South Africa	Morris et al., 1999 ^[205]
De vine®	Phytophthora citrophthora p.v. pal mivora	Strangler vine	USA	
CASST TM	Alternaria cassiae	Sicklepod, senna	Brazil	Charudattan et al., 1982 ^[30]
Smolder®	Alternaria sp.	Dodder	USA	Bewick et al., 2000 [206]
ABG-5003	Cercospora rodmanii	Water hyacinth	USA	Charudattan, 1991, 2001 [31-32]
Dr. BioSedge®	Puccinia canaliculata	Yellow nutsedge	USA	-
Woad Warrior	Puccinia thlaspeos	Isastis tinctoria	USA	-
Camperico®	Xanthomonas Campestris	Annual bluegrass	Japan	-
MycoTech ^{тм}	Chondrostereum purpureum	Various broad leaved trees	Canada;USA	-
Chontrol TM Ecoclear TM	Chondrostereum purpureum	Alders and Hard wood	USA	Barton et al., 2005 [19]
Sarritor	Sclerotinia minor	Dandelion	Australia	Abu-Dieyeh and Watson 2009 [207]

Characteristics of Bioherbicides

Bio-herbicides in culture produce durable and plentiful inoculae. They are cultivated by fermentation for obtaining large masses or for obtaining large yields that are active up to a few days

- Bioherbicides are living innoculum of plant pathogen mainly fungi since the potential of bacteria is hardly explored and viruses proves difficult to handle on the ground of their host specificity and dependence on vectors.
- They are capable of in-vitro culturing in artificial media and mass production.
- They are applied directly to the target weed to kill or reduce the population and growth and they are commercially formulated and spray like hebicides over crops and weeds in the field.

Advantages and Disadvantages of Bioherbicides

The increasing public interest in safe and environmentally friendly products has led to the emergence of numerous new options for pest control, including weed management. Bioherbicides, derived from plant extracts, phytopathogenic microorganisms, or microbial phytotoxins (known as mycoherbicides), represent a valuable approach to weed control (Lamberth & Cai, 2016; Boyetchko, 2004) ^[115, 23]. These bioherbicides offer a sustainable and ecologically sound alternative to conventional synthetic herbicides. They are often biodegradable, with lower toxicity levels, and can

be part of integrated weed management strategies that minimize environmental impact. However, their effectiveness can vary depending on factors such as weather conditions, target weed species, and application methods. Ongoing research and development are essential to improve the efficacy and broaden the spectrum of bioherbicides available for sustainable weed management practices. Bioherbicides typically lack persistent characteristics, meaning they do not remain active in the environment for extended periods. This characteristic reduces the likelihood of soil and water contamination and minimizes adverse effects on non-target organisms. Bioherbicides derived from allelochemicals are particularly benign to both ecosystems and human health (Soltys, 2013) ^[173]. Additionally, some allelochemicals are water-soluble, simplifying their application without the need for surfactants (Dayan, 2009; Vyvvan, 2002) ^[44, 192]. This combination of traits underscores the potential of bioherbicides as a safe and sustainable alternative for weed management, aligning with the growing demand for environmentally friendly solutions. However, further research is necessary to optimize their effectiveness and application methods, ensuring they can be integrated seamlessly into sustainable agriculture practices. Allelochemicals are characterized by their environmentally friendly chemical structures, contrasting with the often complex and potentially harmful structures of synthetic herbicides. Bioherbicides derived from allelochemicals typically exhibit short-lived environmental persistence and

low toxicity, making them safer for the environment and human health (Bailey, 2014) ^[16]. Moreover, these bioherbicides often employ multiple modes of action, which can help reduce the risk of herbicide resistance in weed populations.

Given these characteristics, allelochemicals are considered excellent candidates for the development of bioherbicides, as well as antimicrobial agents and growth regulators. Their natural origins and environmentally friendly attributes make them attractive for various agricultural applications, aligning with the growing demand for sustainable and eco-friendly solutions.

Despite their numerous benefits in sustainable weed control, bioherbicides have limitations that can make them less suitable than synthetic herbicides, especially at the field scale. One significant drawback is their relatively short environmental half-life. While this characteristic reduces environmental toxicity, an effective herbicide needs to persist long enough to achieve the desired effect on weed species (Manahan, 2017)^[119].

Another challenge is the variability in the quantity and content of secondary metabolites produced by plants from the same area or taxonomic group. This variability means that not all plants will exude the same amount or quality of allelochemicals, potentially affecting the consistency and efficacy of bioherbicides (Cheema & Imatomi, 2013; Albert, 2012) ^[39, 5].

Additionally, many allelochemicals are structurally complex and costly to produce, limiting their practicality as agrochemicals. The expense associated with these compounds can hinder their widespread adoption in agriculture despite their potential benefits (Cheema & Imatomi, 2013; Albert, 2012) ^[39, 5]. For example, the cyclic tetrapeptide toxin is an excellent herbicide, but it is very expensive (Imatomi, 2013) ^[69]. Some natural products that exhibit high phytotoxicity can also be highly poisonous to mammals. For example, AAL-toxins are known to be fairly toxic to mammalian cells (Charudattan, 2001) ^[31-32]. Due to these characteristics, the interest in developing these natural phytotoxins into herbicides for weed management has been reduced.

Currently Marketed Products

Since their introduction in 1980, bioherbicides have been part of a broader category of biopesticides that also includes bioinsecticides, biobactericides, biofungicides, and bionematicides. While several biopesticides have been introduced globally, bioherbicides constitute less than 10% of the total market share (Hintz, 2007)^[64].

Most registered bioherbicides are derived from microorganisms (Zeng, 2020) ^[200]. By 2012, seven bioherbicides were registered in the USA, six in Canada, and one each in Ukraine and Japan (Bailey). In 2016, there were thirteen bioherbicides marketed globally, with nine derived from fungal microorganisms, three from bacterial microorganisms, and only one from plant extracts. By 2020, six commercial bioherbicides derived from essential oils and/or their compounds were registered and available in the USA (Verdeguer *et al.* 2020) ^[190].

Approaches to biological control of weeds Classical/Inoculative Approach

Classical / Inoculative approach involves the release of bioagents (insects, fungi, nematodes, fish and other biological system) just for once in the belief that it will readily adapt to the prevailing climate and multiply enough to keep pace with the multiplication rate of weed. The main principle of classical approach is "The regulation of population of an individual pest below the economic threshold". Therefore, biological control exercise to limit their infestation in such a density/level at which they will not be highly damaging to the crops (Jha *et al.*, 2008; Jha *et al.*, 2011; Pahade *et al.*, 2023; Kumhar *et al.*, 2022; Kumbhare *et al.*, 2023; Sahu *et al.*, 2023 and Patidar *et al.*, 2023) ^[87, 81, 135, 111, 113, 109, 151, 153, 139]

Augmentative/Inundative Approach

Several inocula such as fungi, bacteria, parasitic nematodes even viruses having tested control ability over a weed species may be applied by innundative approach.certain philosophy like "Myco-herbicides/bioherbicides is spayed like herbicides does not match with insects, which are released but not sprayed. Therefore, better "bio-herbicides" should include only microorganisms e.g. fungi, bacteria, nematodes viruses but not insects for which classical / Inoculative approach is the main domain.

Comparison between Inoculative and Inundative approach

Economic risk is a critical factor influencing the acceptance of a biocontrol program among farmers and consumers. Farmers, in particular, tend to demonstrate risk-averse behavior, seeking to minimize wide fluctuations in income from year to year (Auld and Tisdell, 1987)^[14]. This risk consideration is more pertinent to the inundative approach than the classical approach in biocontrol.

The classical biocontrol approach typically involves the release of controlled and monitored biocontrol agents by government departments or regulatory bodies. This level of oversight and management helps mitigate economic risks associated with the introduction of biocontrol agents. In contrast, the inundative approach involves the release of large numbers of biocontrol agents, which can lead to greater uncertainty and potential economic risks for farmers. Consequently, farmers may be more cautious in adopting inundative biocontrol methods, preferring the more regulated and controlled classical approach for its predictability and lower economic risk. Bioherbicides differ from classical biocontrol agents in that consumers, such as farmers, are responsible for evaluating their economic benefits compared to chemical alternatives (if available). This evaluation necessitates farmer education programs provided by advisory and extension workers, as well as effective marketing strategies from the manufacturers and distributors of bioherbicides.

The importance of host specificity differs sharply between the classical and inundative approaches. Classical biological control insists upon specificity. The U.S. Environmental Protection Agency (EPA), along with equivalent agencies in other countries like the Australian Plant Quarantine section of the Department of Primary Industries and Energy, imposes stringent and extensive host-range tests on classical biocontrol agents. These tests are designed to ensure that these agents do not attack non-target plants before they are deemed safe for importation and release (Charudattan, 1982) ^[30].

Despite these precautions, there is still cause for concern because the behavior of an exotic pathogen or parasite is challenging to predict accurately (Evans, 1986) ^[53]. This unpredictability was demonstrated by the rapid adaptation of *Puccinia xanthii* Schaw. to both *Helianthus annuus* and *Calendula officinalis* in Australia following an accidental introduction in 1975 (Alcorn, 1976) ^[6]. Prior screening trials in Europe had not indicated that sunflowers were susceptible to this rust, and indeed, sunflowers had been continuously exposed to the fungus in America without issue.

Formulation of bioherbicides

The formulation of bioherbicide active substances is a critical factor in determining their efficacy and success in weed control. Bioherbicides based on microbial metabolites and natural products must be formulated to protect their chemical nature and enhance their entry into plants. Therefore, formulations need to be developed to optimize deployment and efficacy in the field (Morra *et al.*, 2018; Hynes and Boyetchko, 2006) ^[124, 67].

Traditionally, formulation development has focused on adapting ingredients from the chemical pesticide industry, often without considering compatibility with microorganisms or natural products. Successful formulation development has been achieved through collaboration with industries such as food processing and pharmaceuticals, leading to formulations compatible with bioherbicide substances (Hynes and Boyetchko, 2006) ^[67].

The primary functions of bioherbicide formulations should include preparing the weed for infection by the pathogen and protecting the pathogen against environmental constraints while promoting disease development (Charudattan, 2001)^[31-32]. Foliar and stem fungal pathogens, for example, require specific humidity (dew periods) and temperature ranges for full effectiveness. Unique formulations have been developed for mycoherbicides to ensure efficacy after agent delivery in the field. Factors critical in bioherbicide formulation technology, as outlined by Boyetchko et al. (1998) [24], include maintaining or enhancing the effectiveness of the biocontrol agent while ensuring compatibility with conventional field application practices.

Popular formulations for bioherbicides include various emulsions, organosilicon surfactants, hydrophilic polymers, and encapsulated granules made from alginate, starch, or cellulose. Each formulation type has its advantages and disadvantages in promoting the virulence and efficacy of biotic agents and in terms of ease of application (Charudattan, 2001; Hallett, 2005)^[31-32].

Bio herbicides in integrated weed management

Integrated Weed Management (IWM) is a comprehensive and long-term approach to weed control that utilizes all available strategies, including tillage, cultural practices, herbicides, allelopathy, and biological control (Sinodiya and Jha (2014) ^[169], Jha and Kewat (2013) ^[79-80], Tiwari *et al.* (2013) ^[185], and Tiwari *et al.* (2011a) ^[186] Kewat *et al.*, 2009) ^[96]. The goal of IWM is to reduce the weed seed bank in the soil, prevent weed emergence, and minimize competition from weeds growing alongside desired plants (Aldrich and Kremer, 1997) ^[97].

Similar to chemical herbicides, bioherbicides are often most effective when used as part of an integrated approach rather than as a standalone tactic. This integrated approach offers the most promising scenario for bioherbicides to be a practical management option in cropping systems. When viewed as a three-part system, weed management provides several opportunities for integrating bioherbicides at critical stages during weed development: targeting seeds in the soil, controlling growing and competitive plants, and managing seed production (Aldrich and Kremer, 1997)^[97].

Integrating bioherbicides with chemical herbicides

Integrating bioherbicides into weed management strategies can enhance overall control, especially when dealing with multiple weed species. Since most biological control agents are specific to single weed species and fields are often infested with several predominant weeds, combining bioherbicides for one species with herbicides for others makes sense (Kumar *et al.*, 2023, Shri *et al.*, 2014 Jha and Soni 2011 and Sanodiya *et al.* 2013, Kantwa *et al.* 2019, Jha *et al.* 2007 and Kumbhare *et al.*, 2023) ^{[105, 107, 163, 81, 159, 91, 88, 109].}

For instance, the bioherbicide *Fusarium solani f. sp. cucurbitae*, which targets Texas gourd (*Cucurbita texana*), a problematic weed in soybean and cotton fields in the southern United States, was found to be compatible with trifluralin. This compatibility allows for integration into a broader weed management strategy within the crop (Weidemann and Templeton, 1988)^[193].

Moreover, combining reduced rates of herbicides with mycoherbicides can improve the effectiveness of the latter against weeds. For example, *Phoma proboscis* was more effective in controlling field bindweed (*Convolvulus arvensis*) when combined with sublethal doses of 2,4-D compared to when applied alone (Heiny, 1994) ^[63]. Similarly, applying a sublethal dose of the pathogen *Pyricularia setariae* along with one-fourth the rate of three herbicides achieved complete control of green foxtail, demonstrating a synergistic effect between the pathogen and herbicides (Peng and Byer, 2005) ^[140-141].

another example, the fungus In Colletotrichum gloeosporioides f. sp. malvae, which naturally controls round-leaved mallow, achieved adequate control (about 75% kill) when used alone as a bioherbicide (Grant et al., 1990) ^[58]. However, since several chemical herbicides are effective only on round-leaved mallow at the early seedling stage, tank mixtures of the fungus with metribuzin or imazethapyr at recommended rates significantly improved control and reduced biomass production compared to using the fungus or herbicide alone. These examples underscore the effectiveness of combining different weed control strategies and highlight the need for integrated approaches in managing weed infestations.

Integrating bio herbicides with cultural practices

Cultural practices are adaptable for the delivery and integration of bioherbicides in cropping systems. Practices such as crop rotation, tillage, organic amendments, reduced chemical inputs, and crop varietal selection can encourage the development of specific inhibitory microorganisms in the soil and on roots. This can be achieved by enhancing indigenous disease-suppressive rhizobacteria (DRB) or by applying selected cultures along with primary and cover crops.

Combined management activities in long-term cropping systems can select and stimulate soil organisms that directly affect weed growth, leading to high populations of deleterious soil microorganisms that contribute to natural weed suppression (Kremer and Li, 2003) ^[102]. Higher proportions of indigenous rhizobacteria inhibitory to downy brome and jointed goatgrass were detected under either conventional or reduced tillage compared to no-till, suggesting that application of selected DRB during tillage may be effective in integrated weed management (Kremer and Kennedy, 1996) ^[101].

Lindquist *et al.* (1995) ^[116] reported that a natural population of the fungal pathogen *Verticillium sp.* significantly suppressed velvet leaf growth in soybeans under reduced tillage. Addition of composted swine manure to soil inhibited germination and seedling emergence of three weed species, possibly by enhancing soilborne weed-suppressive microorganisms (Menalled *et al.*, 2005) ^[121].

Selectinz highly competitive and allelopathic soybean varieties, matched with compatible bioherbicides, may provide early-season weed suppression and require only minimal subsequent postemergence weed control (Rose *et al.*, 1984)^[105].

Cover crops and mulches are vital for improving soil health and restoring soil productivity in agricultural management systems. They are well-suited for integrating bioherbicides by delivering the agents on seeds and promoting their establishment in soils for an attack on weed seeds and seedlings before planting the main crop. At planting, several cover crop species inoculated with a DRB bioherbicide maintained DRB rhizosphere populations, which transferred to and promoted root colonization of giant foxtail seedlings that emerged later with the primary crop after cover crop termination (Kremer, 2000)^[99]. The combined effects of DRB and allelopathic cover crop residues suppressed weed growth. Formulated bioherbicides are applied at planting to attack weed seeds and seedlings by delivering microbial agents to the soil through direct inoculation of crop seeds or promoting crop root colonization (Skipper *et al.*, 1996)^[170]. Crop roots not only deliver microbial agents to adjacent weed roots but may also maintain or even enhance biotic agent density for seedling attack later in the season. Combining bioherbicides with other control methods can enhance weed control effectiveness. For example, the efficacy of a bioherbicide on hemp sesbania (Sesbania exaltata) was increased by combining selected bacteria with the fungal pathogen Colletotrichum truncatum (Schisler et al., 1991) ^[160]. Similarly, combining a Collectrichum sp. bioherbicide with a naturally occurring rust fungus allowed the bioherbicide to infect the weed host (Xanthium sp.) through rust lesions, leading to the plant's death (Morin et al., 1993)^[123].

Furthermore, combining a seed-feeding insect with seedattacking fungi significantly reduced velvet leaf seed viability and seedling emergence while increasing seed infection compared to using either the insect or fungus alone (Kremer and Spencer, 1989) ^[103]. This approach can effectively manipulate and reduce weed seed banks in soil.

Another strategy involves using soil-applied detrimental bacteria and/or fungi in combination with root-feeding insects, which has been shown to accelerate the growth decline of leafy spurge and knapweed weeds under continuous insect larvae attack (Caesar, 2005) ^[27]. These examples demonstrate how synergistic interactions between bioherbicides and other control methods can enhance weed management efforts.

Synergism between bio-herbicides and chemical herbicides

Acifluorfen and bentazon were the most effective synergists and provided significant control in several weed/pathogen combinations.

Achievements, Developments and Future Challenges

Recent developments have led to over 22 different bioherbicide formulations being registered for weed control, with the global bioherbicide market expected to reach USD 4.14 billion by 2024. However, barriers to their widespread adoption remain. Environmental conditions such as humidity, soil type, temperature, and UV light influence bioherbicide efficacy and formulation. Climate change may also impact weed population dynamics and herbicide resistance, emphasizing the need for improved weed management strategies like bioherbicides.

Formulating and commercializing bioherbicides is challenging due to the need to maintain the viability and stability of living biotic agents. Costs associated with formulation and registration, as well as regulatory hurdles, also hinder their adoption. Overcoming these barriers requires increased education about bioherbicides, technological advancements, and collaboration between governments and agencies to address uncertainties and promote their long-term use. Farmers are more likely to adopt low-cost solutions for weed management, highlighting the importance of addressing economic factors in the adoption of sustainable weed management practices.

Conclusion

It seems like we've summarized the key points quite effectively. Bio-herbicides offer a targeted and environmentally friendly approach to weed management, and integrating them into existing practices can enhance overall effectiveness. Overcoming challenges such as cost and availability will require a concerted effort from various stakeholders, including governments, research institutions, and agricultural producers. Education and technological advancements will play crucial roles in promoting the longterm use of bio-herbicides.

References

- 1. Adetunji CO, Oloke JK, Bello OM, Pradeep M, Jolly RS. Isolation, structural elucidation and bioherbicidal activity of an eco-friendly bioactive 2-(Hydroxymethyl) Phenol, from *Pseudomonas aeruginosa* (C1501) and its ecotoxicological evaluation on soil. Environmental Technology & Innovation. 2019;13:304-317.
- 2. Adetunji CO, Oloke JK, Osemwegie OO. Environmental fate and effects of granular pesta formulation from strains of *Pseudomonas aeruginosa* C1501 and *Lasiodiplodia pseudotheobromae* (C1136) on soil activity and weeds. Chemosphere. 2018;195:98-107.
- 3. Agrawal KK, Bhadauria UPS, Jha A, Jain S. Crop weather relationship studies on chickpea for improving crop adaptation to climate change. Indian Journal of Tropical Agriculture. 2010;28:239-242.
- 4. Aktar W, Sengupta D, Chowdhury A. Impact of pesticides use in agriculture: Their benefits and hazards. Interdisciplinary Toxicology. 2009;2:1-12.

- Albert A. Selective Toxicity: The Physico-Chemical Basis of Therapy. Berlin/Heidelberg, Germany: Springer Science & Business Media; 2012. p. 240-246.
- 6. Alcorn JL. Host range of *Puccinia xanthii*. Transactions of the British Mycological Society. 1976;66:365-367.
- 7. Aldrich RJ, Kremer RJ. Principles in Weed Management. Iowa State University; c1997.
- Algandaby MM, Salama M. Management of the noxious weed *Medicago polymorpha*, L. via allelopathy of some medicinal plants from Taif region, Saudi Arabia. Saudi Journal of Biological Sciences. 2018;25:1339-1347.
- Aliki HM, Reade JP, Back MA. Effects of concentrations of *Brassica napus* (L.) water extracts on the germination and growth of weed species. Allelopathy Journal. 2014;34:287-298.
- Amri I, Hanana M, Jamoussi B, Hamrouni L. Essential oils of *Pinus nigra* JF Arnold Subsp. *Laricio Maire*: Chemical composition and study of their herbicidal potential. Arabian Journal of Chemistry; c2017. p. 10-S3882.
- 11. Andolfi A, Boari A, Evidente M, Cimmino A, Vurro M, Ash G, *et al.* Gulypyrones A and B and Phomentrioloxins B and C produced by *Diaporthe gulyae*, a potential mycoherbicide for saffron thistle (*Carthamus lanatus*). Journal of Natural Products. 2015;78:623-629.
- Aneja KR, Khan SA, Aneja A. Bioherbicides: Strategies, Challenges and Prospects. In: Satyanarayana T, Deshmukh S, Johri B, editors. Developments in Fungal Biology and Applied Mycology. Berlin, Germany: Springer; c2017. p. 449-470.
- Atole A, Gontia AS, Jha A, Upadhyay AS, Nayak PS. Evaluation of maize genotypes for physiological efficiency and productivity under agroclimatic conditions of Kymore plateau zone, Madhya Pradesh. JNKVV Research Journal. 2014;48(1):47-54.
- 14. Auld BA, Tisdell CA. Economic thresholds and response to uncertainty in weed control. Agricultural Systems. 1987;25:219-227.
- 15. Auld BA, Hetherington SD, Smith HE. Advances in bioherbicide formulation. Weed Biology and Management. 2003;3:61-67.
- Bailey KL. The bioherbicide approach to weed control using plant pathogens. In: Integrated Pest Management. Cambridge, MA, USA: Academic Press; 2014. p. 245-266.
- Barfa M, Chouhan M, Verma NS, Lodhi BS, Jha AK. Response of Kabuli Chickpea (*Cicer kabulinum* L.) Varieties to Seed Inoculation with Biofertilizers and Supplementation with Molybdenum. International Journal of Environment and Climate Change. 2023;13(11):3222-3229.
- Barros S, Pedrosa JLF, Gonçalves DR, de Medeiros FCL, Carvalho GR, Gonçalves AH, *et al.* Herbicides of biological origin: A review. Journal of Horticultural Science and Biotechnology. 2021;96:1-9.
- Barton J. Bioherbicides: All in a day's work... for a superhero: What's New in Biological Control of Weeds. Manaaki Whenua, Landcare Research New Zealand Ltd.; c2005. p. 4-6.
- 20. Bhalse L, Jha AK, Verma B, Raghuvanshi S, Porwal M, Sahu MP. Efficacy of pyroxasulfone and its combination against weeds in wheat (*Triticum*)

aestivum). Indian Journal of Agronomy. 2023;68(4):443-446.

- 21. Bhawana S, Girish J, Jha AK, Pratik S. Efficacy of chlorimuron for controlling weeds in soybean. Indian Journal of Weed Science. 2015;48(1):86-89.
- 22. Bowers RC. Commercialization of CollegoTM -an industrialist's view. Weed Science. 1986;34(Suppl):24-25.
- 23. Boyetchko S, Peng G. Challenges and strategies for development of mycoherbicides. In: Fungal Biotechnology in Agriculture, Food, and Environmental Applications; c2004. p. 111-121.
- Boyetchko S, Pederson E, Punja Z, Reddy M. Formulations of biopesticides. In: Boyetchko S, Peng G. Challenges and strategies for development of mycoherbicides. In: Fungal Biotechnology in Agricultural, Food, and Environmental Applications; c1998. p. 111-121.
- 25. Boyette CD. Unrefined corn oil improves the mycoherbicidal activity of *Colletotrichum truncatum* for hemp sesbania (*Sesbania exaltata*) control. Weed Technology. 1994;8:526-529.
- Boyette CD, Hoagland RE, Weaver MA, Stetina K. Biological Control Potential of *Colletotrichum* gloeosporioides for Coffee Senna (*Cassia occidentalis*). American Journal of Plant Science. 2012;3(4):430-436. doi:10.4236/ajps.2012.34052.
- Caesar AJ. Melding ecology, classical weed biocontrol, and plant microbial ecology can improve practices for controlling invasive plant species. Biological Control. 2005;6:240-246.
- 28. Cai X, Gu M. Bioherbicides in organic horticulture. Horticulturae. 2016;2(2):3.
- 29. Carbonari CA, *et al.* Bialaphos and Glufosinate: Bioherbicides from *Streptomyces viridochromogenes*. Journal of Agricultural and Food Chemistry. 2016;64(19):3765-3774.
- Charudattan R. Regulation of microbial weed control agents. In: Charudattan R, Walker HL, editors. Biological Control of Weeds with Plant Pathogens. New York: John Wiley and Sons; c1982. p. 175-188.
- Charudattan R. Biological control of weeds by means of plant pathogens: Significance for integrated weed management in modern agroecology. Biocontrol. 2001;46:229-260.
- 32. Charudattan R. Biological control of weeds by means of plant pathogens: significance for integrated weed management in modern agro-ecology. BioControl. 2001;46:229-260.
- 33. Chauhan A, Jha G, Chourasiya A, Jha A, Joshi J. Effect of tillage and weed management practices and growth productivity and energy analysis of late sown chickpea. International Journal of Agriculture Sciences. 2017;9(5):3779-3781.
- 34. Chauhan A, Chourasiya A, Singh P, Jha A. Effect of different tillage and weed management practices on growth and yield of chickpea. Journal of Plant Development Sciences. 2019;11(5):273-279.
- 35. Chauhan A, Jha G, Chourasiya A, Jha A. Effect of tillage and weed management practices on soil microbial. Journal of Pharmacognosy. 2018;7(1S):1106-1108.

- 36. Chauhan PS, Jha AK, Soni M. Efficacy of *chlorimuron-ethyl* against weeds in transplanted rice. Indian Journal of Weed Science. 2013;45(2):135-136.
- 37. Chauhan BS, Matloob A, Mahajan G, Aslam F, Florentine SK, Jha P. Emerging challenges and opportunities for education and research in weed science. Frontiers in Plant Science. 2017;8:1537-1550.
- 38. Chauhan BS, Matloob A, Mahajan G, Aslam F, Florentine SK, Jha P. Emerging challenges and opportunities for education and research in weed science. Frontiers in Plant Science. 2017;8:1537-1550.
- Cheema ZA, Farooq M, Khaliq A. Application of allelopathy in crop production: Success story from Pakistan. In: Allelopathy. Berlin/Heidelberg: Springer; 2013. p. 113-43.
- 40. Cheng L, Zhu HX, Wei YH, Guo LZ, Weng H, Guo QY. Biological control of Qinghai plateau terrestrial weeds with the *A. alternata* HL-1. Journal of Plant Diseases and Protection. 2021;128:1691-1704.
- 41. Cheng L, Zhu HX, Wei YH, Guo LZ, Weng H, Guo QY. Biological control of Qinghai plateau terrestrial weeds with the *A. alternata* HL-1. Journal of Plant Diseases and Protection. 2021;128:1691-1704.
- 42. Cimmino A, Zonno MC, Andolfi A, Troise C, Motta A, Vurro M, *et al. Agropyrenol*, a phytotoxic fungal metabolite, and its derivatives: A structure-activity relationship study. Journal of Agricultural and Food Chemistry. 2013;61:1779-1783.
- 43. Daniel JJ Jr, Zabot GL, Tres MV, Harakava R, Kuhn RC, Mazutti MA. *Fusarium fujikuroi*: A novel source of metabolites with herbicidal activity. Biocatalysis and Agricultural Biotechnology. 2018;14:314-320.
- Dayan FE, Cantrell CL, Duke SO. Natural products in crop protection. Bioorganic & Medicinal Chemistry. 2009;17:4022-4034.
- 45. DiTomaso JM, van Steenwyk RA, Nowierski RM, Meyerson LA, Doering OC, Lane E, *et al.* Addressing the needs for improving classical biological control programs in the USA. Biological Control. 2017;106:35-39.
- 46. Dubey JK, Kushwaha HS, Jha AK. Growth and biomass productivity of maize as influenced by maize cultivars and maize based cropping system. The Pharma Innovation Journal. 2022;SP-11(7):749-751.
- 47. Dubey JK, Kushwaha HS, Jha AK. Yield and economics of maize under maize-based cropping system in Kymore Plateau of Madhya Pradesh. Annals of Agricultural Research New Series. 2022;43(4):409-412.
- 48. Duke SO, Pan Z, Bajsa-Hirshel J, Boyette CD. The potential future roles of natural compounds and microbial bioherbicides in weed management in crops. Advances in Weed Science. 2022;40:020210054.
- 49. Duke SO, Dayan F, Romagni J, Rimando A. Natural products as sources of herbicides: Current status and future trends. Weed Research. 2000;40:99-111.
- 50. Duke SO, Dayan F, Romagni J, Rimando A. Natural products as sources of herbicides: Current status and future trends. Weed Research. 2000;40:99-111.
- 51. Dwivedi BS, Pandey AK, Tiwari RK, Jha AK, Khamparia NK. Performance of integrated nutrient management on yield and uptake of direct seeded rice. Progressive Agriculture. 2012;12(2):381-385.

- 52. Dwivedi AP, Mishra A, Singh SRK, Singh RP, Jha A. Multiplier effect of zero tillage technology on resource conservation in wheat cultivation. Journal of Community Mobilization and Sustainable Development. 2012;7(1):137-140.
- 53. Evans HC. Biological control of weeds using fungal pathogens. In: Fundamental and Applied Aspects of Invertebrate Pathology, 4th International Colloquium of Invertebrate Biology, Wageningen. 1986. p. 475-480.
- 54. Gangwar S, Naik KR, Jha A, Bajpai A. Soil properties as influenced by organic nutrient management practices under rice based cropping systems. Research on Crops. 2016;17(1):8-12.
- 55. Gautam AK, Shrivastava AK, Jha A. Design parameters of tractor drawn pressurized aqueous fertilizer drill. AMA-Agricultural Mechanization in Asia, Africa and Latin America. 2021;52(3):54-60.
- Gharde Y, Singh PK, Dubey RP, Gupta PK. Assessment of yield and economic losses in agriculture due to weeds in India. Crop Protection. 2018;107:12-18.
- 57. Grand View Research Inc. GVR. Bioherbicides Market Size Worth \$4.14 Billion by 2024 [Internet]. Available from: https://www.grandviewresearch.com/pressrelease/global-bioherbicides-market (accessed on 1 January 1970).
- Grant NT, Prusinkiewicz E, Mortensen K, Makowski RMD. Herbicide interactions with *Colletotrichum* gloeosporioides f. sp. malvae, a bioherbicide for round leaved mallow (*Malva pusilla*) control. Weed Technology. 1990;4:716-723.
- 59. Hall FR, Menn JJ, editors. Biopesticides: Use and Delivery, Methods in Biotechnology; c2002.
- 60. Hallet SG. Where are the bioherbicides? Weed Science. 2005;53:404-415.
- 61. Harding DP, Raizada MN. Controlling weeds with fungi, bacteria, and viruses: A review. Frontiers in Plant Science. 2015;6:659.
- 62. Hasan M, Mokhtar AS, Rosli AM, Hamdan H, Motmainna M, Ahmad-Hamdani MS. Weed control efficacy and crop-weed selectivity of a new bioherbicide Weed Lock. Agronomy. 2021;11:1488.
- 63. Heiny DK. Field survival of *Phoma proboscis* and synergism with herbicides for control of field bindweed. Plant Disease. 1994;78:1156-1164.
- 64. Hintz W. Development of *Chondrostereum purpureum* as a mycoherbicide for deciduous brush control. In: Biological Control: A Global Perspective. CAB International; c2007. p. 284-90.
- 65. Hoagland RE, Boyette CD, Weaver MA, Abbas HK. Bioherbicides: Research and risks. Toxin Reviews. 2007;26:313-342.
- 66. Hosni K, Hassen I, Sebei H, Casabianca H. Secondary metabolites from *Chrysanthemum coronarium* (Garland) flower heads: Chemical composition and biological activities. Industrial Crops and Products. 2013;44:263-271.
- 67. Hynes RK, Boyetchko SM. Research initiatives in the art and science of biopesticide formulations. Soil Biology and Biochemistry. 2006;38:845-849.
- 68. Ibrahim GH. Biological Control of Ryegrass in Wheat Fields by New Isolate of *Urocystis agropyri*. Plant Pathology Research Institute, ARC, Giza, Egypt. 2007;6:260-265.

- 69. Imatomi M, Novaes P, Gualtieri SCJ. Interspecific variation in the allelopathic potential of the family *Myrtaceae*. Acta Botanica Brasilica. 2013;27:54-61.
- 70. Inman RE. A preliminary evaluation of *Rumex* rust as a biological control agent for curly dock. Phytopathology. 1971;61:102-107.
- 71. Jain S, Jha AK. Effect of different tillage systems on yield and economics of mustard varieties and soil properties for Kymore Plateau and Satpura Hillzone of Madhya Pradesh. Journal of Research on Crops. 2012;13(1).
- 72. Jha AK, Shrivastava A, Mehta AK, Billaiya SK, Raghuvanshi NS. Effect of different concentrations of seaweed saps on quality, green fodder, and seed yields of *Berseem (Trifolium alexandrium)*. International Journal of Bio-resource and Stress Management. 2016;27(5):1008-1011.
- 73. Jha AK, Dwivedi BS, Shrivastava AK, Soni M, Taniwaki K, Kokuryu T, *et al.* Effect of drainage, tillage, and land configurations on root, nodules, and yield of soybean in Vertisols. Soybean Research. 2014;12(1, Special Issue):194-98.
- 74. Jha AK, Pandey S, Pandey D, Meshram RK. Effect of fertilizer management with growth regulator on growth and yield of wheat. Pharma Innovation Journal. 2022;11(2):2999-3000.
- 75. Jha AK, Yadav PS, Shrivastava A, Dubey A, Jaiswal A. Influence of tillage and nutrient management on fodder productivity and economics of oat in a rice-oat cropping system. International Journal of Research in Agronomy. 2024;7(4):607-614.
- 76. Jha AK, Shrivastava A. Influence of physiological parameters and yield by different maize-based cropping systems. The Bioscan. 2015;10(1):203-205.
- 77. Jha AK, Shrivastava A, Mehta AK, Billaiya SK, Raghuvanshi NS. Effect of different concentrations of seaweed saps on quality, green fodder, and seed yields of *Berseem (Trifolium alexandrium)*. International Journal of Bio-resource and Stress Management. 2016;27(5):1008-1011.
- 78. Jha AK, Shrivastava A, Raghuvansi NS, Kantwa SR. Effect of weed control practices on fodder and seed productivity of *Berseem* in Kymore plateau and Satpura hill zone of Madhya Pradesh. Range Management and Agroforestry. 2014;35(1):61-65.
- 79. Jha AK, Srivastava A, Raghuvanshi NS, Singh AK. Effect of nitrogen levels on fodder yield and quality of pearl millet genotypes under irrigated condition of Madhya Pradesh. JNKVV Research Journal. 2013;47(2):165-68.
- Jha A, Kewat ML. Weed compositions and seed bank as affected by different tillage and crop establishment techniques in the rice-wheat system. Indian Journal of Weed Science. 2013;45(1):19-24.
- 81. Jha AK, Kewat ML, Upadhyay, Vishwakarma SK. Effect of tillage and sowing management on productivity, economics, and energetics of rice-wheat cropping system for Kamore Plateau and Satpura hill zone of Madhya Pradesh. Indian Journal of Agronomy. 2011;56(1):35-50.
- 82. Jha SK, Sharma AK, Jha AK. Influence of sowing dates and time of fertilizer application on nutrient uptake in soybean crop for Malawa region of Madhya Pradesh. JNKVV Research Journal. 2010;44(2):143-145.

- Jha AK, Soni M. Weed management by sowing methods and herbicides in soybean. Indian Journal of Weed Science. 2013;45(4):250-252.
- 84. Jha AK, Shrivastava A, Nayak S. Integrated nutrient management in forage-based cropping system (*Jowar* + *Cowpea Berseem*). International Journal of Pure and Applied Bioscience. 2018;6(5):1206-1211.
- 85. Jha AK, Yadav PS, Shrivastava A, Upadhyay AK, Sekhawat LS, Verma B, *et al.* Effect of nutrient management practices on productivity of perennial grasses under high moisture condition. AMA, Agricultural Mechanization in Asia, Africa and Latin America. 2023;54(3):12283-12288.
- 86. Jha AK, Shrivastava A, Raghuvanshi NS. Effect of different phosphorus levels on growth, fodder yield, and economics of various cowpea genotypes under Kymore plateau and Satpura hills zone of Madhya Pradesh. International Journal of Agricultural Sciences. 2014;10:409-411.
- 87. Jha AK, Kewat ML, Chaturvedi PL, Sharma RS, Vishwakarma SK. Effect of varying tillage and sowing methods on weed dynamics under rice-wheat cropping system in Kymore plateau and Satpura hill zone of Madhya Pradesh. Indian Journal of Weed Science. 2008;40(1 & 2):37-40.
- 88. Jha AK, Sharma RS, Vishwakarma SK. Development of resource conservation techniques for tillage and sowing management in rice-wheat cropping system under irrigated production system of Kymore Plateau and Satpura hill zone of Madhya Pradesh. JNKVV Research Journal. 2007;41(1):26-31.
- 89. Kaab SB, Rebey IB, Hanafi M, Hammi KM, Smaoui A, Fauconnier ML, *et al.* Screening of Tunisian plant extracts for herbicidal activity and formulation of a bioherbicide based on *Cynara cardunculus*. South African Journal of Botany. 2020;128:67-76.
- Kalam S, Khan NA, Singh J. A novel phytotoxic phenolic compound from *Phoma herbarum* FGCC# 54 with herbicidal potential. Chemistry of Natural Compounds. 2014;50:644-647.
- 91. Kantwa SR, Agrawal RK, Jha A, Pathan SH, Patil SD, Choudhary M. Effect of different herbicides on weed control efficiency, fodder and seed yields of berseem (*Trifolium alexandrinum* L.) in central India. Range Management and Agroforestry. 2019;40(2):323-328.
- 92. Kato-Noguchi H, Kimura F, Ohno O, Suenaga K. Involvement of allelopathy in inhibition of understory growth in red pine forests. Journal of Plant Physiology. 2017;218:66-73.
- 93. Kazinczi G, Lukacs D, Takacs A, Horvath J, Gaborjanyi R, Nadasy M, *et al.* Biological decline of *Solanum nigrum* due to virus infections. Journal of Plant Disease Protection. 2006;32:325-330.
- 94. Kennedy AC. Selective soil bacteria to manage downy brome, jointed goatgrass, and medusahead and do no harm to other biota. Biological Control. 2018;123:18-27.
- 95. Kewat ML, Jha AK, Nayak S. Efficacy of combi against weeds in non-cropped land and microbial properties of soil. Journal of Tropical Forestry. 2011;27(2):72-78.
- 96. Kewat ML, Meena, Vasudev, Sharma Neetu, Jha AK. Effect of time application on the efficacy of combi and glyphosate against *Para grass (Brachiaria mutica)* in

Non-cropped area. Indian Journal of Weed Science. 2009;40(3&4):159-161.

- 97. Koodkaew I, Senaphan C, Sengseang N, Suwanwong S. Characterization of phytochemical profile and phytotoxic activity of *Mimosa pigra* L. Agriculture and Natural Resources. 2018;52:162-168.
- 98. Kotambari Laxmi Bhimappa, Gontia AS, Upadhyaya Anubha, Jha Amit, Singh Ompal. Influence of seed priming on morpho-physiological traits, growth, drought indices, and productivity of maize. International Journal of Current Microbiology Applied Science. 2017;6(9).
- 99. Kremer RJ. Growth suppression of annual weeds by deleterious rhizobacteria integrated with cover crops. In: Spencer NR, editor. Proceedings of Xth International Symposium of Biological Control of Weeds, 4-14 July 1999, Bozeman, MT. Montana State Univ., Bozeman; c2000. p. 931-940.
- 100.Kremer RJ. Bioherbicides and Nanotechnology: Current Status and Future Trends. In: Nano-Biopesticides Today and Future Perspectives. Academic Press; c2019. p. 353-366.
- 101.Kremer RJ, Kennedy AC. Rhizobacteria as biocontrol agents of weeds. Weed Technology. 1996;10:601-609.
- 102.Kremer RJ, Li J. Developing weed suppressive soils through improved soil quality management. Soil Tillage Research. 2003;72:193-202.
- 103.Kremer RJ, Spencer NR. Impact of a seed-feeding insect and microorganisms on velvetleaf (*Abutilon theophrasti*) seed viability. Weed Science. 1989;37:211-216.
- 104.Kumar D, Singh M, Kumar S, Meena RK, Yadav MR, Makarana G, et al. Productivity and Quality Enhancement in Fodder Maize (*Zea mays*) Cultivars through Nutrient Management Strategies. Indian Journal of Agricultural Sciences. 2022;92:126-130.
- 105.Kumar B, Shaloo, Bisht H, Meena MC, Dey A, Dass A, et al. Nitrogen management sensor optimization, yield, economics, and nitrogen use efficiency of different wheat cultivars under varying nitrogen levels. Frontiers in Sustainable Food Systems. 2023;7:1228221.
- 106.Kumar R, Bohra JS, Kumawat N, Kumar A, Kumari A, Singh AK. Root growth, productivity and profitability of baby corn (*Zea mays* L.) as influenced by nutrition levels under irrigated eco-system. Research Crops. 2016;17:41-46.
- 107.Kumar S, Sanodiya P, Jha AK, Sahu MP, Verma B. Effect of 2,4-D sodium salt on weeds, growth and yields in rabi maize (*Zea mays* L.). Indian Journal of Agronomy. 2023;68(4):100-103.
- 108.Kumar Vinod, Shukla RS, Moham mad Hussain Ekram, Pandey Suneeta, Jha Amit Kumar. Phenotypic Diversity of Newly Developed Cytoplasmic Lines of Wheat Based on Multivariate Analysis. Frontiers in Crop Improvement. 2021;9:3336-3344. Print ISSN: 2393-8234.
- 109.Kumbhare Rahul, Jha AK, Patel Rajendra, Ajay Jaiswal, Gupta Vikas, Kumar Pramod, *et al.* Optimum dose of fertilizer with growth regulators for higher yield of wheat (*Triticum aestivum* L.). The Pharma Innovation Journal. 2023;12(11):1117-1123.
- 110.Kumbhare Rahul, Jha Amit Kumar, Anjana Gopilal, Patel Raghav, Tekam Yagini. Combination of Fertilizer and Growth Regulators Impact on Nutrient Balance in

Wheat Crop (*Triticum aestivum* L.). International Journal of Plant & Soil Science. 2023;35(22):823-832.

- 111.Kumhar Bheru Lal, Agrwal KK, Jha AK, Rai HK, Kumar Vijay, Choudhary Mukesh. Productivity and economical viability of grass-based cropping systems. Range Management and Agroforestry. 2022;43(1):167-171.
- 112.Kumhar Bheru Lal, KK Agrawal, Jha AK. Weed screening in grass based cropping system in Central India. Journal of Biotechnology and Crop Science. 2020;9(14):94-99.
- 113.Kumhar Bheru Lal, KK Agrawal, AK Jha, Rai HK. Variability of soil infiltration rate in different grass based cropping system in Kymore Plateau Satpura Hills region. Forage Res. 2021;47(3):325-328.
- 114.Kautariye JK, Sharma JK, Kewat ML, Jha AK, Dubey Jitendra. Efficacy of Different Herbicides Against Weeds and Economics of the Treatment in Direct Seeded Rice Kymore Plateau Region of Madhya Pradesh. Trends in Biosciences. 2017;10(2):636-639.
- 115.Lamberth C. Naturally occurring amino acid derivatives with herbicidal, fungicidal or insecticidal activity. Amino Acids. 2016;48:929-940.
- 116.Lindquist JL, Maxwell BD, Buhler DD, Gunsolus JL. Velvetleaf (*Abutilon theophrasti*) recruitment, survival, seed production and interference in soybean (*Glycine max*). Weed Science. 1995;43:226-232.
- 117.Lodhi Bhumika Singh, Kumar Pramod, Chouhan Monika, Rajpoot Alok, Jha Amit. Comprehensive Genetic Analysis of Yield and Yield-Related Traits in Soybean Germplasms for Enhanced Crop Improvement. International Journal of Plant & Soil Science. 2023;35(22):9-17. Article no.IJPSS.108908 ISSN: 2320-7035
- 118. Malviya P, Jha AK, Upadhyay VB. Effect of different proportions of vermicompost and fertilizers on growth and yield of scented rice and soil properties. Annals of Agricultural Research New Series. 2012;33(4):228-234.
- 119. Manahan SE. Environmental Chemistry. CRC Press; c2017.
- 120.Matzrafi M, Seiwert B, Reemtsma T, Rubin B, Peleg Z. Climate change increases the risk of herbicide-resistant weeds due to enhanced detoxification. Planta. 2016;244:1217-1227.
- 121.Menalled FD, Buhler DD, Liebman M. Composted swine manure effects on germination and early growth of crop and weed species under greenhouse conditions. Weed Technology. 2005;19:784-789.
- 122. Monika Raghuwanshi, Amit Kumar Jha, Kratika Naik, Anamika Pandey. Influence of weed management strategies on nutrient uptake and soil properties in fodder maize cultivation. International Journal of Research in Agronomy. 2024;7(5):24-29.
- 123.Morin L, Auld BA, Brown JF. Synergy between Puccinia xanthii and Colletotrichum orbiculare on Xanthium occidentale. Biological Control. 1993;3:296-310.
- 124. Morra MJ, Popova IE, Boydston RA. Bioherbicidal activity of *Sinapis alba* seed meal extracts. Industrial Crops Production. 2018;115:174-181.
- 125. Mortensen K. Effects of *Colletotrichum* gloeosporioides f. sp. malvae on plant development and biomass of non-target crops under field conditions. Weed Research. 1997;37:351-360.

- 126.Mortensen K. Biological control of weeds using microorganisms. In: Boland GJ, Kuykendal LD, eds. Plant-Microbe Interaction and Biological Control. Marcel Dekker; c1998. p. 223-248.
- 127. Motmainna M, Juraimi AS, Uddin M, Asib NB, Islam AKM, Hasan M. Bioherbicidal properties of *Parthenium hysterophorus, Cleome rutidosperma*, and *Borreria alata* extracts on selected crop and weed species. Agronomy. 2021;11:643.
- 128.MP Sahu, ML Kewat, AK Jha, RK Tiwari, VK Choudhary, Vikash Singh, *et al.* Effect of weed control and crop residue mulches on weeds and yield of chickpea in Kymore plateau and Satpura hills of Madhya Pradesh. The Pharma Innovation Journal. 2023;12(12):242-245.
- 129.Nichols V, Verhulst N, Cox R, Govaerts B. Weed dynamics and conservation agriculture principles: A review. Field Crops Research. 2015;183:56-68.
- 130.Oehrens E. Biological control of blackberry through the introduction of the rust, *Phragmidium violaceum*, in Chile. FAO Plant Protection Bulletin. 1977;25:26-28.
- 131.Oerke EC. Crop losses to pests. Journal of Agricultural Science. 2006;144:31-43.
- 132.Ootani MA, dos Reis MR, Cangussu ASR, Capone A, Fidelis RR, Oliveira W, *et al.* Phytotoxic effects of essential oils in controlling weed species *Digitaria horizontalis* and *Cenchrus echinatus*. Biocatalysis, Agriculture and Biotechnology. 2017;12:59-65.
- 133.Osadebe VO, Dauda N, Ede AE, Chimdi GO, Echezona BC. The use of bioherbicides in weed control: Constraints and prospects. African Journal of Agricultural Technology. 2021;21:37-54.
- 134. Pacanoski Z. Bioherbicides. In: Herbicides, Physiology of Action, and Safety. Intech Open; 2015. pp. 253-274.
- 135.Pahade S, Jha AK, Verma B, Meshram RK, Toppo O, Shrivastava A. Efficacy of sulfentrazone 39.6% and pendimethalin as a pre emergence application against weed spectrum of soybean (*Glycine max* L. Merrill). International Journal of Plant & Soil Science. 2023;35(12):51-58.
- 136.Parmar S, Jha AK, Dubey J. Efficiency of different post emergence herbicides on weeds and economics of the treatments, yield and productivity in different soybean varieties. Eco. Env. & Cons. 2017;23:146-150.
- 137.Patel R, Jha AK, Verma B, Porwal M, Toppo O, Raghuwanshi S. Performance of pinoxaden herbicide against complex weed flora in wheat (*Triticum aestivum* L.). International Journal of Environment and Climate Change. 2023;13(7):339-345.
- 138.Patidar J, Kewat ML, Jha AK. Present status of weed flora in soybean crop. The Pharma Innovation. 2019;8(7):717-720.
- 139.Patidar J, Kewat ML, Sondhiya S, Jha AK, Gupta V. Bio-efficacy of fomesafen + fluazifop-p-butyl mixture against weeds and its effect on productivity and profitability of soybean (*Glycine max*) in Central India. Indian Journal of Agricultural Sciences. 2023;93(7):750-755.
- 140.Peng G, Byer KN. Interactions of *Pyricularia setariae* with herbicides for control of green foxtail (*Setaria viridis*). Weed Technology. 2005;19:589-598.
- 141.Peng G, Byer KN. Interactions of *Pyricularia setariae* with herbicides for control of green foxtail (*Setaria viridis*). Weed Technology. 2005;19:589-598.

- 142.Piyaboon O, Pawongrat R, Unartngam J, Chinawong S, Unartngam A. Pathogenicity, host range and activities of a secondary metabolite and enzyme from *Myrothecium roridum* on water hyacinth from Thailand. Weed Biology and Management. 2016;16:132-144.
- 143.Porwal M, Verma B, Jha AK. New and innovative technologies and machinery in conservation agriculture. Climate smart agriculture. Principles and practices; c2023. p. 113-127. ISBN: 978-81-19149-15-5
- 144.Raghav P, Jha AK, Badal V, Rahul K, Richa S. Bioefficacy of pinoxaden as post-emergence herbicide against weeds in wheat crop. Pollution research. 2023;42(1):115-117.
- 145.Raghuwanshi M, Jha AK, Verma B, Yadav PS, Shrivastava A. Weed dynamics of fodder maize as influenced by different herbicides. International Journal of Environment and Climate Change. 2023a;13(7):245-251.
- 146.Raghuwanshi M, Jha AK, Verma B, Sahu MP, Dubey A. Assessing the effect of weed management practices on weed flora, growth and yield of fodder maize (*Zea mays* L.). International Journal of Plant & Soil Science. 2023b;35(11):112-120.
- 147.Rao AN. Weed Science in the Tropics: Principles and Practices. Oxford and IBH Publishing Co. Pvt. Ltd.; c2000.
- 148.Reinhart KO, Carlson CH, Feris KP, Germino MJ, Jandreau CJ, Lazarus BE, *et al.* Weed-suppressive bacteria fail to control *Bromus tectorum* under field conditions. Rangeland Ecology & Management. 2020;73:760-765.
- 149.Rheinhold L, Simon HM. Weed control by rhizobacteria: Plant pathogens. Weed Technology. 2000;10:610-620.
- 150.Rose SJ, Burnside OC, Specht JC, Swisher BA. Competition and allelopathy between soybeans and weeds. Agronomy Journal. 1984;76:523-528.
- 151.Sahu MP, Kewat ML, Jha AK, Choudhary VK, Verma B, Patidar J, *et al.* Effect of crop residue and weed management on weed incidence, soil moisture and yield of chickpea. Indian Journal of Agronomy. 2023;68(4):404-412.
- 152.Sahu MP, Kewat ML, Jha AK, Sondhia S, Choudhary VK, Jain N, *et al.* Weed prevalence, root nodulation and chickpea productivity influenced by weed management and crop residue mulch. AMA, Agricultural Mechanization in Asia, Africa and Latin America. 2022;53(6):8511-8521.
- 153.Sahu V, Kewat ML, Verma B, Singh R, Jha AK, Sahu MP, *et al.* Effect of carfentrazone-ethyl on weed flora, growth and productivity in wheat. The Pharma Innovation Journal. 2023;12(3):3621-3624.
- 154.Sairam G, Jha AK, Verma B, Porwal M, Dubey A, Meshram RK. Effect of mesotrione 40% SC on weed growth, yield and economics of maize (*Zea mays* L.). International Journal of Environment and Climate Change. 2023;13(7):608-616.
- 155.Sairam G, Jha AK, Verma B, Porwal M, Dubey A, Meshram RK. Effect of mesotrione 40% SC on weed growth, yield and economics of maize (*Zea mays* L.). International Journal of Environment and Climate Change. 2023;13(7):608-616.
- 156.Sairam G, Jha AK, Verma B, Porwal M, Sahu MP, Meshram RK. Effect of pre and post-emergence

herbicides on weed flora of maize. International Journal of Plant & Soil Science. 2023;35(11):68-76.

- 157.Sairam G, Jha AK, Verma B, Porwal M, Sahu MP, Meshram RK. Effect of pre and post-emergence herbicides on weed flora of maize. International Journal of Plant & Soil Science. 2023;35(11):68-76.
- 158.Salikram Mohare, Tiwari G, Tripathi N, Rawat A, Jha A, Ramakrishnan RS, *et al.* Comparative Efficacy of Chemical and Plant-based Herbicides on Weed Management in Winter Sown Ashwagandha Crop (*Withania somnifera* (L.) Dunal). Biological Forum -An International Journal. 2023;15(2):444-449.
- 159.Sanodiya P, Jha AK, Shrivastava A. Effect of integrated weed management on seed yield of fodder maize. Indian Journal of Weed Science. 2013;45(3):214-216.
- 160. Schisler DA, Howard KM, Bothast RJ. Enhancement of disease caused by *Colletotrichum truncatum* in *Sesbania exaltata* by coinoculating with epiphytic bacteria. Biological Control. 1991;1:261-268.
- 161.Sekhar JC, Sandhya S, Vinod KR, Banji D, Sudhakar K, Chaitanya RSN AKK. Plant toxins-useful and harmful effects. Hygeia Journal of Drugs & Medicine. 2012;4:79-90.
- 162.Sharma A, Agrawal KK, Sharma JK, Jha AK. Bioefficacy of bentazone 48% SL as post-emergence against weeds in direct-seeded rice. Indian Journal of Weed Science. 2020;52(1):74-77.
- 163.Shri Sonam, Jha AK, Shrivastava A. Evaluation of different intercropping systems for maximum productivity and economics in maize. Annals of Agricultural Research, New Series. 2014;35(2):201-205.
- 164.Singh H, Jha G, Babu S, Jha AK. Effect of seed rate and sowing depth on growth, yield attributes and yield of irrigated wheat (*Triticum aestivaum*) in Madhya Pradesh. Indian Journal of Agronomy. 2013a;58(2):57-59.
- 165.Singh V, Agrawal KK, Jha AK, Sahu MP. Effect of forchlorfenuron on yield and economics of transplanted rice. Ind J Pure App Biosci. 2019;7(4):411-414.
- 166.Singh Vikash, Agrawal KK, Jha AK. Effect of forchlorfenuron on yield and economics of transplanted rice. Progressive Agriculture. 2019;2:290-292.
- 167.Singh H, Jha G, Rawat A, Babu S, Jha AK. Low seed rate at surface sowing enhances resilience of physiological parameters and economics of wheat (*Triticum aestivum*). The Indian Journal of Agricultural Sciences. 2013b;83(8).
- 168.Singh S, Chhokar RS, Gopal R, Ladha JK, Gupta RK, Kumar V, *et al.* Integrated weed management: A key to success for direct-seeded rice in the Indo-Gangetic Plains. In Integrated Crop and Resource Management in the Rice—Wheat System of South Asia. International Rice Research Institute. c2009. p. 261-278
- 169.Sinodiya P, Jha AK. Effect of weed management control practices on nutrient uptake and soil properties in fodder maize. JNKVV Research Journal. 2014;48(1):60-63.
- 170.Skipper HD, Ogg AG Jr, Kennedy AC. Root biology of grasses and ecology of rhizobacteria for biological control. Weed Technology. 1996;10:610-620.
- 171.Smith RJ. Integrating microbial herbicides with current pest management strategies. John Wiley & Sons; c1982.

- 172.Smith RJ. Blending biological control agents with chemical pesticides. In: Microbial Control of Weeds. Chapman and Hall; c1991. p. 189-208.
- 173.Soltys D, Krasuska U, Bogatek R, Gniazdow A. Allelochemicals as bio-herbicides: present status and future prospects. In: Price AJ, Kelton JA, editors. Herbicides—Current Research and Case Studies in Use. In Tech; c2013. p. 517-542.
- 174.Somasundram C, Razali Z, Santhirasegaram V. Reviewing organic food production in Malaysia. Horticulturae. 2016;2:12.
- 175.Soni M, Jain KK, Jha AK. Impact of post-emergence herbicide timing on weed seed bank in transplanted rice (*Oryza sativa* L.) at Kyamore plateau and Satpura hill zones of Madhya Pradesh. Advances in Applied Research. 2013;5(2).
- 176.Soni M, Jain KK, Jha AK. Weed dynamics and yield in transplanted rice (*Oryza sativa* L.) with post-emergence herbicides. Journal of Current Advances in Agricultural Sciences. 2012;4(2):165-167.
- 177.Sun Y, Kaleibar BP, Oveisi M, Müller-Schärer H. Exploring climate change: Lessons from plant invasion science and weed science. Frontiers in Agronomy. 2021;2:1-7.
- 178. Synowiec A, Możdżeń K, Krajewska A, Landi M, Araniti F. *Carum carvi* L. essential oil: A potential botanical herbicide against *Echinochloa crus-galli* (L.) P. Beauv. in maize cultivation. Industrial Crops & Products. 2019;140:111652.
- 179.Nirala T, Jha AK, Verma B, Yadav PS, Anjna M, Bhalse L. Assessing the efficacy of pinoxaden on weed population and wheat yield (*Triticum aestivum* L.). Biological Forum: An International Journal. 2022;14(4):558-561.
- 180.Tataridas A, Kanatas P, Chatzigeorgiou A, Zannopoulos S. Navigating sustainable crop and weed management under the EU Green Deal era: A comprehensive guide. Agronomy. 2022;12:589.
- 181.Templeton GE, Heiny JP. Pioneering biological herbicides. In: Biological Control in Agricultural IPM Systems. 1989. pp. 173-185.
- 182.Tisdell CA, Auld BA, Menz KM. On assessing the value of biological control of weeds. Protection Ecology. 1984;6:169-179.
- 183. Tiwari RK, Mahajan G, Jha A, Singh SK, Tripathi SK. Growth Efficiency, Productivity and Economics of Direct Seeded Rice as Influenced by Nitrogen Level and Weed Management. Journal of Pure and Applied Microbiology. 2017 Jun;11(2):987-991.
- 184. Tiwari RK, Dwived BS, Deshmukh G, Pandey AK, Jha A. Effect of weed control treatments on growth of little seed cannary grass and productivity of wheat. Indian Journal of Weed Science. 2011;43(3&4):239-240.
- 185.Tiwari RK, Jha A, Tripathi SK, Khan IM, Rao SK. Rice based cropping system and climate change. JNKVV Research Journal. 2013;47(1):239-247.
- 186. Tiwari RK, Khan IM, Singh N, Jha A. Chemical weed control in wheat through on form demonstration in Rewa district of Madhya Pradesh. Indian Journal of Weed Science. 2011a;43(3&4):215-216.
- 187.Tomar DS, Jha AK, Porwal M, Verma B, Tirkey S, Khare Y, *et al.* Efficacy of Halauxifen-methyl+ Florasulam against Complex Weeds in Wheat under Kymore Plateau and Satpura Hill Zone of Madhya

Pradesh, India. International Journal of Plant & Soil Science. 2023;35(15):161-171.

- 188. Tomar DS, Jha AK, Verma B, Meshram RK, Porwal M, Chouhan M, *et al.* Comparative Efficacy of Different Herbicidal Combinations on Weed Growth and Yield Attributes of Wheat. International Journal of Environment and Climate Change. 2023;13(8):889-898.
- 189. Toppo O, Kewat ML, Jha AK, Yadav PS, Verma B. Effect of Sowing time and weed management practices on weed dynamics, productivity and economics of direct-seeded rice. Ecology, Environment, and Conservation. 2023;29(3):80-85.
- 190. Verdeguer M, Sánchez-Moreiras AM, Araniti F. Phytotoxic effects and mechanism of action of essential oils and terpenoids. Plants. 2020;9:1571.
- 191.Singh V, Agrawal KK, Jha AK. Effect of Forchlorfenuron on growth and yield of Rice (*Oryza* sativa L.) during Kharif season of Central India. International Journal of Current Microbiology and Applied Sciences. 2019;8(9):2331-2338.
- 192. Vyvyan JR. Allelochemicals as leads for new herbicides and agrochemicals. Tetrahedron. 2002;58:1631-1646.
- 193. Weidemann GJ, Templeton GE. Control of Texas gourd, *Cucurbita texana*, with *Fusarium solani* f. sp. *cucrbitae*. Weed Technology. 1988;2:271-274.
- 194. Willer H, Yussefi M. The World of Organic Agriculture 2005—Statistics and Emerging Trends. International Federation of Organic Agriculture Movements. 2015.
- 195.Xu X, et al. Colletotrichum gloeosporioides Bioherbicide for Controlling Malva pusilla. Phytopathology. 2019;109(3):459-467.
- 196. Yadav PS, Jha AK, Kewat ML, Sahu MP, Verma B. Effect of sowing management and herbicides application on weeds and yield of berseem (*Trifolium alexandrinu*). Indian Journal of Agronomy. 2024;69(1):71-77.
- 197. Yadav PS, Jha AK, Pachauri V, Verma B, Shrivastava A, Chouhan M, *et al.* Oat genotypes response to different nitrogen levels under agro-climatic condition of Kymore Plateau zone of Madhya Pradesh. Pharma Innovation. 2023a;12(4):2371-2374.
- 198. Yadav PS, Kewat ML, Jha AK, Hemalatha K, Verma B. Effect of sowing management and herbicides on the weed dynamics of berseem (*Trifolium alexandrinum*). Pharma Innovation. 2023b;12(2):2845-2848.
- 199. Yadav PS, Kewat ML, Jha AK, Sahu MP, Verma B, Toppo O. Floristic composition of weeds as influence by sowing time and herbicides in berseem. Ecology, Environment, and Conservation. 2023c;29(3):64-68.
- 200.Zeng P. Bio-Herbicides: Global Development Status and Product Inventory; c2020. Available online: http://news.agropages.com/News/NewsDetail---34164.htm (accessed on 21 February 2021).
- 201.Zhang Y, Yang X, Zhu Y, Li L, Zhang Y, Li J, et al. Biological control of Solidago canadensis using a bioherbicide isolate of Sclerotium rolfsii SC64 increased the biodiversity in invaded habitats. Biological Control. 2019;139:104093.
- 202.Zimdahl RL. Fundamentals of Weed Science. Academic Press; c2018.
- 203.Zimdahl RL (Ed.). Integrated Weed Control for Sustainable Agriculture. Burleigh Dodds Publishing; c2018.

- 204.Ziska LH. Climate change and the herbicide paradigm: Visiting the future. Agronomy. 2020;10:1953.
- 205.Morris, Michael L, Cheryl R. Doss. How does gender affect the adoption of agricultural innovations? The case of improved maize technology in Ghana; c1999.
- 206. Irving AJ, Coutts AA, Harvey J, Rae MG, Mackie K, Bewick GS, *et al.* Functional expression of cell surface cannabinoid CB1 receptors on presynaptic inhibitory terminals in cultured rat hippocampal neurons. Neuroscience. 2000 Jun 1;98(2):253-62.
- 207. Abu-Dieyeh MH, Watson AK. Increasing the efficacy and extending the effective application period of a granular turf bioherbicide by covering with jute fabric. Weed technology. 2009 Dec;23(4):524-530.