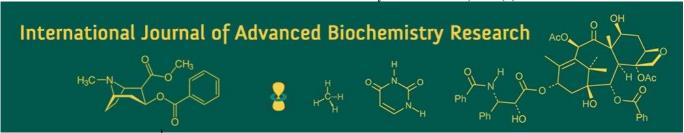
International Journal of Advanced Biochemistry Research 2024; SP-8(5): 184-187



ISSN Print: 2617-4693 ISSN Online: 2617-4707 IJABR 2024; SP-8(5): 184-187 www.biochemjournal.com Received: 21-02-2024 Accepted: 24-03-2024

Lavanya Killi

Ph.D Scholar, Veterinary Surgery and Radiology, College of Veterinary Science and AH, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, India

Indramani Nath

Professor and Head, Department of Veterinary Surgery and Radiology, College of Veterinary Science and AH, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, India

Anjan Kumar Sahoo

Assistant Professor, Veterinary Surgery and Radiology, College of Veterinary Science and AH, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, India

Susen Kumar Panda

Professor and Head, Department of Veterinary Pathology, College of Veterinary Science and AH, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, India

Bijay Kumar Patra

Associate Professor, Veterinary Clinical Complex, College of Veterinary Science and AH, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, India

Subhranshu Sekhar Biswal

Assistant Professor, Animal Reproduction, Gynaecology and Obstetrics, College of Veterinary Science and AH, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, India

Corresponding Author: Anjan Kumar Sahoo

Assistant Professor, Veterinary Surgery and Radiology, College of Veterinary Science and AH, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, India

Cancer stem cell and epithelial mesenchymal transition in chemoresistance of canine solid tumours

Lavanya Killi, Indramani Nath, Anjan Kumar Sahoo, Susen kumar Panda, Bijay Kumar Patra and Subhranshu Sekhar Biswal

DOI: https://doi.org/10.33545/26174693.2024.v8.i5Sc.1151

Abstract

There is increasing evidence for the presence of cancer stem cells in several solid tumors, and these cancer stem cells have a potential role in tumor initiation, aggression, and recurrence. Solid tumors occur frequently in dogs, where such tumors exhibit complexity when examined histologically. These tumors are composed not only of proliferative luminal epithelial cells, but also of myoepithelial cells and /or mesenchymal cells with cartilage and osseous tissues in a solitary mass. The origin of the complexed histogenesis remains speculative, but cancer stem cells (CSCs) are likely involved. Canine CSCs are relatively resistant to the cytotoxic effects of common chemotherapeutic drugs and ionizing radiation, indicating that failure of clinical therapy to eradicate canine mammary cancer may be due to the survival of CSCs. The epithelia to mesenchymal transition (EMT) has been associated with cancer invasion, metastasis, and the acquisition of stem cell characteristics. Present investigation shows that canine CSCs predominantly express mesenchymal markers and are more invasive than parental cells, indicating that these cells can be induced to undergo EMT by transforming growth factor-β (TGF) and that these cells have an increased ability to form tumor sphere. CSCs possess self-renewing capacity, differentiation potential, high tumorigenicity in immunodeficient mice, and resistance to chemotherapy and radiation. Analysis of the characteristics of CSCs may contribute to the elucidation of the histogenesis underlying canine solid tumors, formulation of novel CSC-targeted therapeutic strategies, and development of biomarkers for early diagnostics and prognostic applications.

Keywords: Solid tumor, tumor growth factor, cancer stem cell, glioblastoma, cluster of differentiation (CD)

Introduction

Solid cancer consists of heterogeneous cells that contain a sub population of tumor cells with stem cell properties, including self-renewal capacity, differentiation potential, tumorigenicity in immunodeficient mice, and resistance to chemotherapy and radiation [1, 2]. Such tumor cells, termed cancer stem cells (CSCs) or tumor-initiation cells (TICs), are generated either from mutational events in normal tissue stem cells or from the acquisition of stem cell properties by differentiated cells; these tumor cells exist at the apex of a hierarchy of cancer tissues [3, 4, 5] in various cancers such as skin, liver, and glioblastoma, mammary carcinoma. CSCs are organized as tree-like hierarchies [6, 7]. CSCs have the capacity to self-renew, recurrence, to initiate and maintain the tumor, can produce heterogeneous lineages of cancer cells to compose the bulk of the tumor and is the primary cause of metastasis [8, 9, 10]. The cancer stem cell model therefore proposes that tumor development is akin to abnormal organogenesis [8]. In addition, CSCs are resistance to many current cancer treatments, including chemo- and radiation therapy. Therefore, conventional therapies, while killing the bulk of the tumor cells, ultimately fail because they do not eliminate the CSC population, which survives to regenerate the tumor [11]. Further understanding the properties and mechanisms of CSCs is essential in the development of effective-anti-cancer therapies. In humans CSCs were first identified in acute myeloid leukemia [12] and more recently in melanomas [13], glioblastomas [14] and epithelial cancers [15]. In the canine model, we were the first to identify CSCs of a canine osteosarcoma cell line [16] and have subsequently isolated CSCs from a range of canine solid tumors including glioma, hemangiosarcoma and squamous cell carcinoma. Recent evidence has suggested that tumor progression metastasis is dependent upon aberrant activation of epithelial to mesenchymal transition (EMT) in

cancer cells, resulting in the acquisition of invasive and metastatic properties [17]. Classically, EMT is an evolutionarily conserved developmental pathway involved in tissue morphogenesis, organ fibrosis and wound healing [18]. The hallmark of EMT is the loss of cell surface E-cadherin, which is associated with disassembly of adherent junctions, acquired motility and expression of mesenchymal markers including Vimentin and Fibronectin [19]. The EMT program is regulated by multiple transcription factors, including Twist, Snail and members of the zinc finger and homeodomains (zhf) family [20]. It is now known that EMT activation is also associated with the maintenance of stem cell properties, and in vitro it has been that emergence of CSCs occurs as a result of EMT [20].

Properties of Cancer stem cells

CSC share similar properties and presence of surface marker with the normal stem cells present in adult mammalian CSC and stem cells have many similar characteristics like as self-renewal, indefinite selfreplication, asymmetric cell division, generating a large number of differentiated cells and expressing specific surface marker and regulatory molecules [21]. Though, both stem cell and CSC exist for a prolonged period of time but the major difference between their functional properties is that normal stem cells always function under control whereas the division and differentiation of CSC are out of control leading to large number of tumor cells with diversified cell population. In a nutshell, CSC is defined by four distinct properties (a)self-renewal—like stem cells, the CSCs sub population in a culture plate are transplanted through multiple generation (b)differentiation— CSCs have the capacity to differentiate into bulk population of daughter tumor cells (CSCs and non-tumor cells (non-CSCs) indicating its pluripotency. (c) tumorigenicity—a small proportion or subpopulation of CSCs have tumorigenic potential when transplanted into SCID/nude mice or animals and (d) CSCs in addition to common surface marker like stem cells, express specific surface markers by which these are separated from non-stem cells [22].

Tumor microenvironment and characterisation of CSCs

Inside a tumor there are diversified population of cells starting from normal fibroblast, cancer associated fibroblast (CAF), normal stem cells, T cell, endothelial cells, cancer progenitor cell, cancer stem cells (CSCs), cells in the epithelial mesenchymal transition (EMT), extracellular matrix (ECM). Since, CSC and stem cells share many common properties, the isolation and identification of CSC can be done with several in vitro assays such as sphere forming assays [23], Hoechst dye exclusion (SP cells) [24], detection of enzymatic activity of aldehyde dehydrogenase1 (ALDH1) [25], detection of surface markers [24], signalling pathway identification [26], serial colony-forming unit assays (replating assays) [27], label retention assays [1], and migration assays [28]. These in vitro assays are used to isolate stem cells in a tumor but in addition serial transplantation in animal models is required to identify the particular CSCs [1]. Still advanced and optimised methods are currently used to isolate and identify the CSCs. Those are Flow cytometry analysis, fluorescent associated cell sorting (FACS) analysis, polymerase chain reaction (PCR) analysis, immunofluorescence staining analysis which uses cell surface specific markers for CSCs such as surface cell-

adhesion molecules (e.g., CD133, CD24, hyaluronic acid (HA) receptor CD44), cytoprotective enzymes (such as aldehyde dehydrogenase, ALDH), transcription factors or stem cell pluripotency marker (e.g., OCT-4, SOX-2), and drug efflux pumps (e.g., ATP-binding cassette (ABC) drug transporters and multidrug resistance transporter1, MDR1) [24]. Among these CD133, CD24 and CD44 surface marker are commonly used. In canine mammary carcinoma, sphere forming assay are used to identify CSCs as free floating spheres are characteristic of the solid tumors in vitro and they are rich in stem cells and progenitors [29]. Currently, the isolation of CSCs in head and neck cancer (HNSCC) involves (1) detection of side-population phenotypes by Hoechst 33342 exclusion, (2) sphere-formation assays, (3) assessment of aldehyde dehydrogenase (ALDH) activity, and (4) identification of CSC-specific cell surface markers CD44, CD133, and ALDH through FACS immunohistochemistry [30].

Immune evasion and cause of metastasis

The two main reasons for progression, spreading and metastasis of cancer are through CSCs and epithelial mesenchymal transition (EMT). While CSCs initiates tumorigenesis, the EMT helps in formation of new CSCs. EMT is a complex cellular event in which epithelial cells lose their morphology or phenotype by shedding cell-to-cell adhesion molecules, such as desmosomes, tight and gap junctions, lose apical-basal polarity and attain frontal-rear polarity and acquire a mesenchymal phenotype that is associated with high motility and invasive properties [31]. Depending upon the process of transformation EMT is classified into Type I, Type II and Type III EMT. Type I is associated with embryogenesis and organogenesis, Type II with wound healing, tissue regeneration and organ fibrosis whereas Type III is mainly associated with tumor progression [32]. Cells undergoing EMT are identified through loss of epithelial marker such as E-cadherin, EpCAM and cytokeratins and gain of mesenchymal markers like Fibronectin, Vimentin and N cadherin. The process invol ves activation of nuclear transcription factors (ZEB1, SNAIL, SLUG, TWIST, KLF8, and E47), signal transduction pathways (HOX transcription factors, the Wnt-Beatenin, Notch and Sonic Hedgehog (SHh) pathways, and the Polycomb gene *Bmi-1*, and TGF-β signaling) [33] and extracellular matrix (ECM). In Pancreatic ductal adenocarcinoma (PDAC), one of the lethal malignancies nestin plays a major role in tumor initiation and maintenance [34]. Both CSC and EMT are responsible for tumor formation, invasion, metastasis and chemoresistance associated with ovarian cancers [35].

Conclusion

As these cells play a major role in cancer initiation and progression, the isolation of these cells is of major interest for the development therapeutic approaches aiming at directed targeting of these cells. In recent years, targeted immunotherapeutic drugs have been developed as for treatment of malignancy and lengthening overall survival times (OST) in canine patients.

References

1. Clarke MF, Dick JE, Dirks PB, Eaves CJ, Jamieson CHM, Jones DL, *et al.* Cancer stem cells-perspectives on current status and future directions: AACR

- Workshop on cancer stem cell. cancer Research. 2006;66:9339-9344.
- 2. Nguyen LV, Vanner R, Dirks P, Eaves CJ. Cancer stem cells: an evolving concept. Nature .Rev. cancer 2012;12:133-143.
- 3. Kreso A, Dick JE. Evolution of the cancer stem cell model. Cell stem cell. 2014;14:275-291.
- Sahoo AK, Nath I, Senapati SB, Panda SK, Das MR, Patra BK. Comparative Evaluation of Nutraceuticals (Curcuma longa L., Syzygium aromaticum L. and Olea europaea) with Single-agent Carboplatin in the Management of Canine Appendicular Osteosarcoma. Indian Journal of Animal Research; c2021. DOI: 10.18805/IJAR.B-4485
- Sahoo AK, Nath I, Senapati SB, Panda SK, Das MR, Patra BK. Apocrine Gland Anal Sac Adenocarcinoma in Dogs: 22 Cases (2015-2020). Indian Journal of Animal Research, 2021. DOI: 10.18805/IJAR.B-4371
- Gasch C, French B, O'Leary JJ, et al. Catching moving targets: cancer stem cell hierarchies, therapy-resistance & considerations for clinical intervention. Mol Cancer. 2017;16:43. https://doi.org/10.1186/s12943-017-0601-3
- Sahu BP, Majee P, Singh RR, Sahoo AK, Nayak D. Comparative analysis, distribution, and characterization of microsatellites in Orf virus genome. Scientific Reports. 2020;10:13852. https://doi.org/10.1038/s41598-020-70634-6
- 8. Pang LY, Argyle DJ. Using naturally occurring tumours in dogs and cats to study telomerase and cancer stem cell biology. Biochimica et Biophysica Acta. 2009;1792:380-391.
- Sethi RK, Senapati SK, Selim AM, Acharya AP, Mishra C, Das M, et al. Molecular epidemiology of lumpy skin disease outbreak in Odisha, India. Veterinary Research Communications. 2022;46(3):711-717.
- 10. Mishra S, Sahu SK, Panigrahi S, Biswal SS, Mishra SR, Ranjan R, *et al.* Comparative therapeutic efficacy of levofloxacin, ornidazole and alpha tocopherol combination with prostaglandin F2α on IL-6 and IL-10 transcript level in longstanding cases of endometritis in crossbreed Jersey cows. Iranian Journal of Veterinary Research. 2018;19(3):217-224.
- 11. Diehn M, Clarke MF. Cancer stem cells and radiotherapy: New insights into tumor radioresistance. J Natl. cancer Inst. 2006;98:1755-1757.
- 12. Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. Nature Medicine. 1997;3:730-737.
- 13. Fang D, Nguyen TK, Leishear K, *et al.* Tumorigenic subpopulation with stem cell properties in melanomas. cancer Research. 2005;65:9328-9337.
- 14. Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, *et al.* Identification of a cancer stem cell in human brain tumors. Cancer Researcch. 2003;63:5821-5828.
- Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. Proc. Natl.Acad. Sci, USA. 2003;100:3983-3988.
- 16. Wilson H, Huelsemeyer M, Chun R, Young KM, Friedrichs K, Argyle DJ. Isolation and Characterization

- of cancer stem cells from canine osteosarcoma. vet J. 2008;175:69-75.
- 17. Theiry JP. Epithelial-mesenchymal transitions in development and pathologies. Current opinion in cancer cell biology. 2003;15:740-74.
- 18. Singh A, Settleman J. EMT, cancer stem cells and drug resistance: An emerging axis of evil in the war on cancer. Oncogene. 2010;29:4741-4751.
- 19. Gupta PB, Onder TT, Jiagn G, Tao K, Kuperwasser C, Weinberg RA, *et al.* Identification of selective inhibitors of cancer stem cells by high-throughput screening. Cell. 2009;138:645-659.
- 20. Mani SA, Guo W, Liao, Mj, Eaton EN, Ayyanan A, Zhou AY, *et al*. The epithelial-mesenchymal transition generates cells with properties of stem cells. Cells. 2008;133:704-715.
- 21. Pannuti A, Foreman K, Rizzo P, Osipo C, Golde T, Osborne B, *et al.* Targeting notch to target cancer stem cells. Clinical Cancer Research. 2010;16:3141-3152.
- 22. Li C, Heidt DG, Dalerba P, Burant CF, Zhang L, Adsay V, *et al.* Identification of pancreatic cancer stem cells. Cancer research. 2007;67(3):1030-1037.
- 23. Ghani FI, Yamazaki H, Iwata S, Okamoto T, Aoe K, Okabe K, *et al.* Identification of cancer stem cell markers in human malignant mesothelioma cells. Biochemical and Biophysical Research Communication. 2011;404:735-742.
- 24. Mannelli G, Gallo O. Cancer stem cells hypothesis and stem cells in head and neck cancers. Cancer Treatment review. 2012;38:515-539.
- 25. Kim RJ, Nam JS. OCT4 expression enhances features of cancer stem cells in a mouse model of breast cancer. Laboratory Animal Research. 2011;27:147-152.
- 26. Elizabeth O-S, Jose' Luis G-M, Elizabeth L, Alejandro G-C. Cancer stem cells in solid tumors, markers and therapy. Stem Cell Human Disease. 2012, 117-148.
- 27. Kitamura H, Okudela K, Yazawa T, Sato H, Shimoyamada H. Cancer stem cell: implications in cancer biology and therapy with special reference to lung cancer. Lung Cancer. 2009;66:275-281.
- 28. Biddle A, Liang X, Gammon L, Fazil B, Harper LJ, Emich H, *et al.* Cancer stem cells in squamous cell carcinoma switch between two distinct phenotypes that are preferentially migra-tory or proliferative. Cancer Research. 2011;71:5317-5326.
- 29. Michishita M, *et al.* Characterization of spheres derived from canine mammary gland adenocarcinoma cell lines. Research in Veterinary Science; c2010. doi:10.1016/j.rvsc.2010.11.016
- 30. Tirino V, Desiderio V, Paino F, De Rosa A, Papaccio F, La Noce M, *et al.* Cancer stem cells in solid tumors: an overview and new approaches for their isolation and characterization. FASEB J: official publication of the Federation of American Societies for Experimental Biology. 2013;27(1):13-24
- 31. Thiery JP, Acloque H, Huang RYJ, Nieto MA. Epithelial-mesenchymal transitions in development and disease. Cell. 2009;139(5):871-890.
- 32. Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. Journal of clinical investigation. 2009;119(6);1420-1428.
- 33. Nurwidya F, Takahashi F, Murakami A, Takahashi K. Epithelial mesenchymal transition in drug resistance

- and metastasis of lung cancer. Cancer Research and Treatment. 2012;44(3):151-156.
- 34. Skoda J, Hermanova M, Loja T, Nemec P, Neradil J, Karasek P, *et al.* Co-Expression of Cancer Stem Cell Markers Corresponds to a Pro-Tumorigenic Expression Profile in Pancreatic Adenocarcinoma. PLoS One. 2016;11(7):e0159255.
 - doi: 10.1371/journal.pone.0159255. PMID: 27414409; PMCID: PMC4945008.
- 35. Suster NK, Virant-Klun I. Presence and role of stem cells in ovarian cancer. World J Stem Cells. 2019;11(7):383-397.