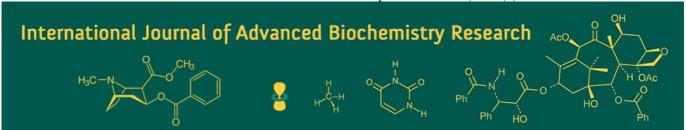
International Journal of Advanced Biochemistry Research 2024; SP-8(4): 547-549



ISSN Print: 2617-4693 ISSN Online: 2617-4707 IJABR 2024; SP-8(4): 547-549 www.biochemjournal.com Received: 02-01-2024 Accepted: 11-03-2024

Soundarya TC

Department of Veterinary Medicine, Veterinary College, Hebbal, Bengaluru, Karnataka, India

Anil Kumar MC

Department of VCC, Veterinary College, Hebbal, Bengaluru, Karnataka, India

Kshama MA

Department of Veterinary Medicine, Veterinary College, Hebbal, Bengaluru, Karnataka, India

Shivaraj Murag

Department of VMC, IAH & VB, Hebbal, Bengaluru, Karnataka, India

Anjan Kumar KR

Department of VPP, Veterinary College, Hassan, Karnataka, India

Pavithra BH

Department of VPT, Veterinary College, Gadag, Karnataka, India

Corresponding Author: Soundarya TC Department of Veterinary Medicine, Veterinary College, Hebbal, Bengaluru, Karnataka, India.

Diagnosis of immune mediated haemolytic anemia in dogs affected with *B. gibsoni*

Soundarya TC, Anil Kumar MC, Kshama MA, Anjan Kumar KR, Shivaraj Murag and Pavithra BH

DOI: https://doi.org/10.33545/26174693.2024.v8.i4Sg.1076

Abstract

The present study was carried out to diagnose immune mediated hemolytic anemia associated with *Babesia gibsoni* in dogs. A total of 136 dogs suspected for *B. gibsoni* infection were examined by microscopy and screened by PCR, out of which 64 dogs were positive for *B. gibsoni* infection. Among 64 dogs, 28 dogs were positive for IMHA by Coombs test, 21 dogs were positive for IMHA by Saline agglutination test and 18 dogs had spherocytosis.

Keywords: Immune mediated haemolytic anemia, Coombs test

Introduction

The present study entitled was carried out to diagnose immune mediated hemolytic anemia associated with *Babesia gibsoni* in dogs. The study was conducted in dogs presented to small animal OPD, Department of Veterinary Medicine with clinical signs of anorexia, pyrexia, lethargy, pale mucous membrane, lymphadenopathy, ascites, vomiting, diarrhea, icterus, hematuria suggestive of canine babesiosis. A total of 136 dogs suspected for *B. gibsoni* infection were examined by microscopy and screened by PCR, out of which 64 dogs were positive for *B. gibsoni* infection. Among 64 dogs, 28 dogs were positive for IMHA by Coombs test, 21 dogs were positive for IMHA by Saline agglutination test and 18 dogs had spherocytosis.

Further these dogs were subjected to saline agglutination test, spherocytosis in blood smear examination and Coombs test for diagnosis of immune mediated haemolytic anaemia (IMHA).

Materials and Methods

Clinical cases

Dogs presented to the Department of Veterinary Medicine, Veterinary College Hospital, Hebbal, Bengaluru with a history of tick infestation, high fever, pale / icteric mucous membrane, inappetence, anaemia, lethargy, vomiting, diarrhoea, hematuria were screened by saline agglutination test, spherocytosis and Coombs test for diagnosing IMHA. These dogs were screened for *B. gibsoni* infection by microscopic examination of blood smears and PCR. Eighteen dogs infected with *B. gibsoni*, positive for saline agglutination test, spherocytosis and Coombs test were selected for the study.

In-saline agglutination test

One drop of EDTA blood was mixed with one to four drops of normal saline on a glass slide. The agglutination of RBCs was assessed macroscopically and then it was observed under the microscope for RBC agglutination which appears like cluster of grapes.

Examination of blood smear for B. gibsoni organisms and spherocytosis

Thin blood smears were prepared from the whole blood sample collected, which was stained with Giemsa stain. Smears were examined under the oil immersion 100X objective of the microscope to detect the presence of *B. gibsoni* organisms and spherocytosis.

Coombs Test

Coombs test was carried out in suspected cases of IMHA in dogs. The collected blood sample was centrifuged at 1500 rpm for 5 minutes at room temperature. 4.9 ml of normal saline was added to 0.1 ml of packed RBCs. The red blood cells were washed 4 times in normal saline and finally 4.9 ml of normal saline was added to make 2% suspension.

Coombs test procedure

100 μL of normal saline was pipetted into 4 test tubes. Two fold dilutions of the Coombs reagent was prepared by Pippeting 100 μL of Coombs reagent (MP biomedicals) to the first tube, mixed and transferred 100 μL from the 1st tube to the 2nd tube, additionally 100 μL of the mixture was transferred from 2nd tube to the 3rd tube. 100 μL was discarded from 3rd tube. The control 4th test tube had only normal saline. Furthermore 100 μL of washed 2% RBCs were added to all the test tubes, gently mixed and was incubated at 37 °C for 30 minutes. After the incubation time was complete, all the four test tubes were centrifuged for 1 minute.

Interpretation

The contents of each tube were evaluated by placing a few drops of solution on a clean glass slide, covered with glass coverslip and examined at a magnification of 100X. Presence of clumps and large aggregates of RBCs was interpreted as positive result of Coombs test while absence of clumping was considered as negative. The clumps should not be present in control cells.

Results and Discussion Diagnosis of IMHA

Out of 64 dogs positive for *B. gibsoni*, 28 dogs showed positive for IMHA by Coombs test, 21 dogs were positive by saline agglutination test and 18 dogs had spherocytosis.

Saline agglutination test

Out of 64 dogs positive for *B. gibsoni*, 21 dogs showed positive for IMHA by saline agglutination test.

Sphericity count

Out of 64 dogs positive for *B. gibsoni*, 18 dogs had spherocytosis.

Coombs test

Out of 64 dogs positive for *B. gibsoni*, 28 dogs showed positive for IMHA by Coombs test accounting to 43.75 percent.

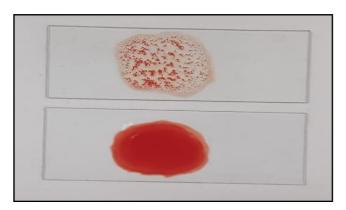


Plate 1: Comparision of positive and negative saline agglutination

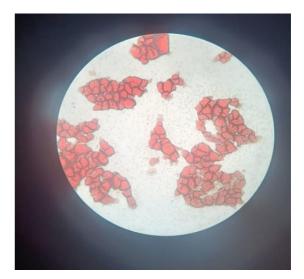


Plate 2: Positive saline agglutination test under the microscope (100X)

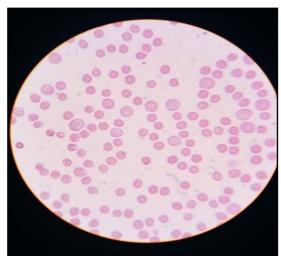


Plate 3: Presence of spherocytes in giemsa stained blood smear

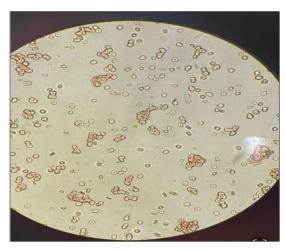


Plate 4: Positive coombs test (40X)

Saline agglutination test

In the present study, 32.812 percent (21 dogs) of dogs were positive for IMHA caused by *B. gibsoni* by saline agglutination test. Weinkle *et al.* (2005) ^[5] reported that 40 to 89 percent of dogs with IMHA had positive saline agglutination test. Lachungpa *et al.* (2020) ^[4] reported that 21 percent of dogs were positive for IMHA by saline agglutination test. The reason for in-saline slide

agglutination was to break up rouleaux formation but not erythrocyte aggregates using saline. This variation in the results could be because a positive saline agglutination is seen when a high level of anti-RBC antibodies are present that are attached to more than one cell.

Spherocytosis

Spherocytosis was observed in 18 dogs (28.125%) with IMHA caused by *B. gibsoni* in the present study. Partial phagocytosis of RBC results in removal of a part of RBC membrane, these RBCs form spherocytes. Spherocytosis is the hallmark of IMHA (Caviezel *et al.* 2014) ^[1]. A spherocyte count of 2+ or greater was highly suggestive of IMHA (Nelson and Couto, 2009) ^[3]. While spherocytosis is recognized as a hallmark of IMHA, accurate evaluation requires meticulous examination and identification due to potential discrepancies among observers which could result in false negative results (Caviezel *et al.* 2014) ^[1].

Coombs test

Out of 64 dogs positive for *B. gibsoni*, 28 dogs (43.75 percent) were positive by Coombs test.

Our study is in accordance with Farewell *et al.* (1982) ^[2] who reported that out of 37 *B. gibsoni* and *B. canis* infected dogs tested, 31 were Coombs test-positive.

Interpretation

Coombs test was more effective in the diagnosis of IMHA as compared to Saline agglutination test or spherocytosis in the blood smear examination.

Conclusion

Study highlights the utility of various diagnostic methods in detecting immune-mediated hemolytic anemia (IMHA) in dogs infected with Babesia gibsoni. The saline agglutination test demonstrated a positivity rate of 32.812%, aligning with previous findings, albeit showing variability likely attributed to differing antibody levels. Spherocytosis, a hallmark of IMHA, was present in 28.125% of cases, emphasizing its diagnostic significance despite potential observer variability. Furthermore, the Coombs test revealed a positivity rate of 43.75%, consistent with prior research. These diagnostic tools collectively contribute to the accurate identification and management of IMHA in Babesia gibsoni-infected dogs, emphasizing the importance of comprehensive evaluation in clinical practice.

References

- 1. Caviezel LL, Raj K, Giger U. Comparison of 4 direct Coombs' test methods with polyclonal antiglobulins in anemic and nonanemic dogs for in-clinic or laboratory use. J Vet. Intern. Med. 2014;28(2):583-591.
- Farwell GE, LeGrand EK, Cobb CC. Clinical observations on Babesia gibsoni and Babesia canis infections in dogs. J Am. Vet. Med. Assoc. 1982;180(5):507-511.
- Nelson RW, Couto CG. Small animal internal medicine. MOSBY Elsevier, S. Louis, Missoui. 2009, 1410
- Lachungpa CG, Chandrasekaran D, Thilagar MB, Kumar TMA. Treatment of secondary immune mediated hemolytic anaemia of dogs in Chennai, Tamil Nadu. Journal of Animal Research. 2020;10(4):535-541.

5. Weinkle TK, Center SA, Randolph JF, Warner KL, Barr SC, Erb HN. Evaluation of prognostic factors, survival rates, and treatment protocols for immunemediated hemolytic anemia in dogs: 151 cases (1993–2002). Journal of the American Veterinary Medical Association. 2005;226(11):1869-1880.